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Organic Preparations and Procedures International

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t902189982>

APPROACHES TO PARTIAL SYNTHESSES OF 11-OXO STEROIDS. A BRIEF REVIEW

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To cite this Article Gao, Hongwu(1997) 'APPROACHES TO PARTIAL SYNTHESSES OF 11-OXO STEROIDS. A BRIEF REVIEW', *Organic Preparations and Procedures International*, 29: 5, 499 – 539

To link to this Article: DOI: 10.1080/00304949709355229

URL: <http://dx.doi.org/10.1080/00304949709355229>

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APPROACHES TO PARTIAL SYNTHESSES OF 11-OXO STEROIDS. A BRIEF REVIEW

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INTRODUCTION

The introduction of an oxo functional group at the 11 β -position of steroids remains an area of interest because it provides the activated site for the further elaboration of steroid rings. Estradiol derivatives of general structure **1** (Fig. 1) encompass a variety of interesting and valuable compounds.

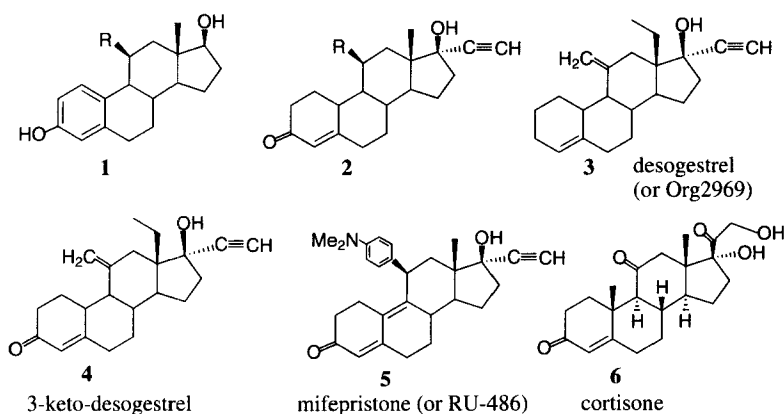


Fig. 1

Those bearing small carbon substituents at the 11 β -position (R = ethyl,¹ chloromethyl,² vinyl³, ethynyl,⁴ ethenyl,⁴ phenyl⁴) are the best known ligands for the estrogen receptor, their affinity being more than 10 times higher than that of estradiol, the natural ligand. Thus, in the synthesis of estradiol derivatives designed as probes for the estrogen receptor, the inclusion of one of such groups (particularly the ethyl and chloromethyl) is often used to secure high binding affinities.^{5,6} In addition to these functions, appropriate 11 β -substitution results in compounds endowed with pure anti-estrogenic activity.⁷ The synthesis of 11 β -substituted estradiol derivatives involves the addition of lithium or Grignard reagents to 11-carbonyl group of an estradiol derivative, followed by the acid-catalyzed dehydration and catalytic hydrogenation (R = ethyl,¹ chloromethyl,² vinyl³) or the stereoselective

deoxygenation of the resulting 11-hydroxy steroids by means of an ionic hydrogenation (R = ethynyl,⁴ ethenyl,⁴ phenyl⁴).

Since some 11-substituted 19-nor-steroids **2** (Fig. 1), particularly 11 β -chloro⁸ and 11 β -methyl⁹ derivatives, were found to be the very biologically active compounds, studies of ovulation inhibitory activity of 11-substituted 19-nor-steroids have been carried out.¹⁰ The differences in biological activity could reasonably be correlated with two steric effects introduced by the 11 β -substituent. These were a change in the overall shape of the 11 β -substituent and the angular methyl group, and direct steric hindrance of the steroid-receptor protein binding. The resulting study¹¹ of 11-alkylidene 19-nor-steroids led a powerful progestogen widely used in oral contraceptives, desogestrel (**3**, Org2969),¹² and its active metabolite-3-keto-desogestrel **4**.¹³ The 11-methylene group could be introduced from 11-oxo-19-nor-steroid in three ways: 1) the addition of methylmagnesium iodide or methyllithium followed by dehydration in formic acid 2) Wittig reaction 3) Peterson olefination.¹⁰

The other stereospecific introduction of 11 β -carbon substituents, such as in mifepristone (**5**, RU-486),^{14, 15} could be completed by the copper-catalyzed conjugate addition of organometallic reagents to vinylloxirane which could be synthesized from the intermediate of 11-oxo-19-nor-steroid.¹⁶

On account of its dramatic influence on chronic rheumatoid arthritis and acute rheumatic fever,¹⁷ cortisone (**6**) is produced commercially. Numerous partial and total syntheses of this compound have been performed by chemical and microbiological methods. A common feature of all of the partial syntheses of cortisone from deoxycholic acid devised by Kendall and co-workers is the multi-step method to shift the oxygen function from C-12 to C-11.¹⁸

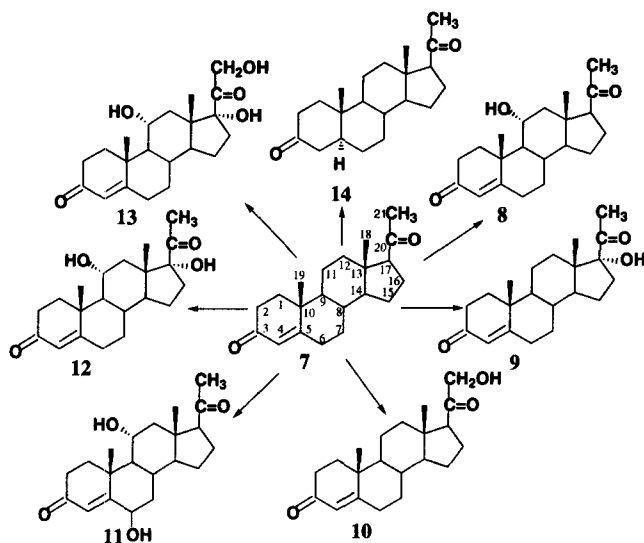
I. SYNTHETIC APPROACHES TO 11-OXO STEROIDS WITH 4-EN-3-ONE

1. Microbiological Transformation

a) Fermentation of Progesterone

11 α -OH-progesterone

Incubation of *Aspergillus ochraceus* on progesterone **7** yielded the corresponding 11 α -hydroxyprogesterone **8** as the main product, together with 6 β ,11 α -dihydroxy derivative **11** (Scheme 1a).¹⁹⁻³⁰ Gaitonde and co-workers²⁵ explored the effect of *N*-methyl-*N*'-nitro-*N*-nitrosoguanidine (NTG) on ability of spores of *Aspergillus ochraceus* to transform progesterone and obtained 85-90% of **8** and 5-10% of **11**. Chopra and Somal²⁶ developed a mutant of *Aspergillus ochraceus* to convert progesterone to 11 α -hydroxyprogesterone in high yield (**8**: 90%; **11**: minimal). The action of *Aspergillus ochraceus* TS on progesterone only gave 11 α -hydroxyprogesterone.³⁰ C-11 Hydroxylation of progesterone with *Aspergillus niger* led to the corresponding 11 α -hydroxyprogesterone along with five by-products (Scheme 1b).³¹⁻³⁹ Paper chromatographic analysis of the mixture indicated that the progesterone was transformed to 11 α -hydroxyprogesterone in 47-62%.³⁵ The other species like *Aspergillus phoenicis*⁴⁰⁻⁴², *Aspergillus fischeri*^{43, 44}, *Aspergillus nidulans*⁴⁵ also could transform progesterone to 11 α -hydroxyprogesterone and other related by-products.



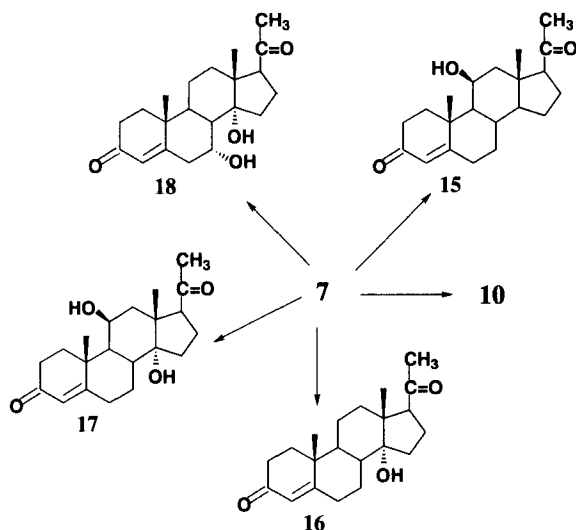
Reagents and Results: a) *Aspergillus ochraceus*: 8 and 11; b) *Aspergillus niger*: 8, 9, 10, 11, 12 and 13; c) *Rhizopus nigricans*: 8, 9, 10, 11 and 14; d) *Rhizopus Arrhizus*: 8, 9, 10 and 11.

Scheme 1

The conversion of progesterone to 11 α -OH-progesterone by *Rhizopus nigricans*, was first reported by Peterson and co-workers in 1952.^{46,47} In this microbiological transformation, 11 α -hydroxyprogesterone is the main product formed (8: 85-95%). In addition, small amounts of a dihydroxyprogesterone (11: 0.5%) and 11 α -hydroxyallopregnane-3,20-dione (14: 0.5-5%) were also isolated (Scheme 1c). The other studies revealed that two additional by-products were formed in this oxidation.⁴⁸⁻⁵⁸ The yield of 6 β ,11 α -dihydroxyprogesterone could be diminished by the addition of 0.02M ascorbic acid to the reaction mixture.⁵³ The nonsterile fermentation of progesterone using a strain of *Rhizopus nigricans* gave a yield of more than 80%.⁴⁹ The rate of 11 α -hydroxylation of progesterone with *Rhizopus nigricans* could be accelerated by glucose.⁵⁷ Peterson's further studies⁴⁷ revealed that *Rhizopus arrhizus* produced a lower yield (50%) of 8 and higher yields (5-15%) of 11. The other two by-products were reported later (Scheme 1d).⁵⁹ Recent investigations showed that supplementation of the medium with critical micelle concentration (CMC) resulted in the production of *Rhizopus arrhizus* mycelium with an increased specific capacity to transform progesterone to 11 α -hydroxyprogesterone.⁶⁰ Special species like *Rhizopus stolonifer*⁶¹, *Penicillium*⁶², *G. Khaki* and *G. Lagenarium*⁶³ could also transform progesterone to 11 α -hydroxyprogesterone with some by-products. Aside from the 11 α -position, the microbial introduction of hydroxy groups into positions 6, 17 and 21 are chemically activated by carbonyl groups.

11 β -OH-progesterone

11 β -Hydroxylation of progesterone by *Curularia lunata* was first described by Kita and Shull.⁶⁴ These workers did not determine the chemical structures of four other transformation products that were produced. In later studies,^{65,66} structural details were provided (Scheme 2). The yields of



Reagents and Results: *Curvularia lunata*, 15, 10, 16, 17 and 18.

Scheme 2

products oxidation at C 11 β - and 21-positions were markedly improved in the presence of Mg²⁺, Fe²⁺, Mn²⁺, Zn²⁺ and Ca²⁺.⁶⁷ C-11 Hydroxylation of 19-norprogesterone with *Curularia lunata* led to the corresponding 11 β -hydroxy analog.⁶⁸

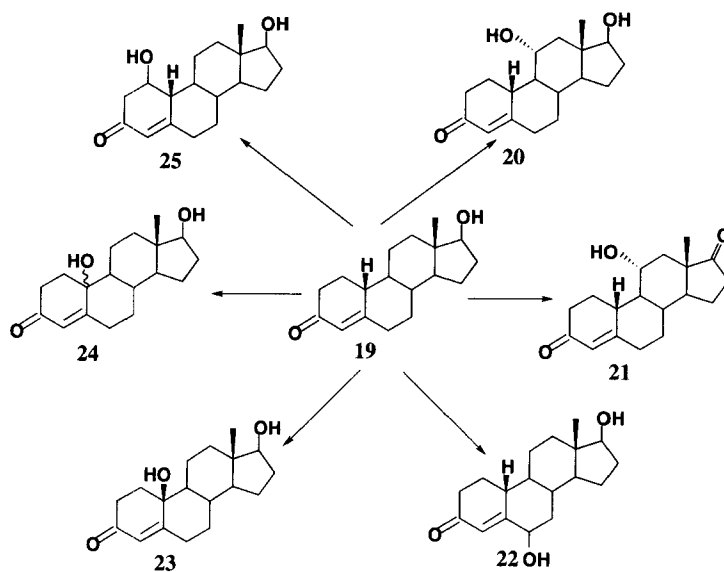
b) Fermentation of 19-Nortestosterone

11 α -OH-19-nortestosterone

In 1956, Peterson and co-workers⁶⁹ reported the preparation of a 10 ξ -hydroxy-19-nortestosterone by the microbiological action of *Rhizopus nigricans* on 19-nortestosterone (Scheme 3a). Hydroxylation proceeded mainly at the 6 β -position (**22**: 18%), while reaction at the 11 α - and 10 ξ -positions (**20**: 4% and **24**: 1.2%) occurred only to a minor extent. Thus, this mold contained an enzyme which could hydroxylate 19-nortestosterone at a tertiary position. *Aspergillus ochraceus* fermentation of racemic 19-nortestosterone yielded *l*-10 β -hydroxy derivative (**23**: 13%), together with *d*-11 α -hydroxy (**20**: 37%), *l*-1 β -hydroxy (**25**: 0.3%), *dl*-6 β -hydroxy (**22**: 0.07%) derivatives (Scheme 3b).⁷⁰ Similarly, the fermentation of *d*-19-nortestosterone with *Aspergillus ochraceus* led to a different product mixture, a 72% yield of 11 α -OH-19-nortestosterone and a small amount of 11 α -hydroxy-estr-4-ene-3,17-dione (**21**: 10%) were obtained (Scheme 3c). In an additional example, the microbiological attack of *Aspergillus ochraceus* on 19-nortestosterone gave the same mixture of products (**20**: 34% and **21**: 20%) (Scheme 3d).⁷¹ Compound **21** (the by-product from the microbiological conversion of 19-nortestosterone with *Aspergillus ochraceus*) can be obtained by fermentation of 19-norandrost-4-ene-3,17-dione with *Aspergillus ochraceus* in a good yield (**21**: 68%) (Scheme 4).⁷¹

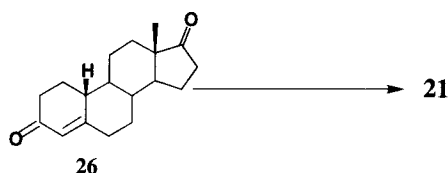
11 β -OH-19-nortestosterone

By fermentation of 19-nortestosterone with *Curvularia lunata*, Szpilfogel and co-workers⁷¹ obtained 10 β -OH-19-nortestosterone as the major product (**23**: 36%) with the 11 β -OH- (**27**: 3.2%),



Reagents and Results: a) *Rhizopus nigricans*, **20**: 4%; **22**: 18%; **24**: 1.2%;
 b) *Aspergillus ochraceus*, *d*-**20**: 37%; *dl*-**22**: 0.07%; *l*-**23**: 13%; *l*-**25**: 0.3%;
 c) *Aspergillus ochraceus*, **20**: 72%; **21**: 10%; d) *Aspergillus ochraceus*, **20**: 34%; **21**: 20%.

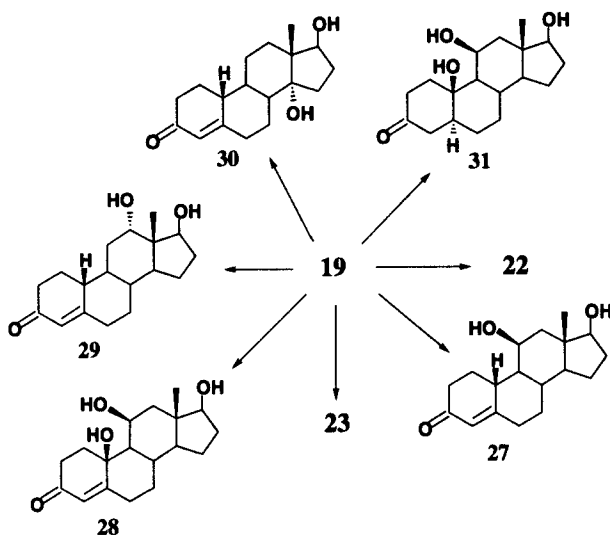
Scheme 3



Reagents and Results: *Aspergillus ochraceus*, **21**: 68%.

Scheme 4

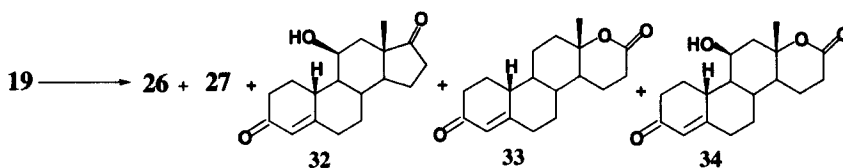
10 β ,11 β -dihydroxy- (**28**: 0.7%) and 14 α -OH-derivatives (**30**: 2.3%) being isolated as minor products (Scheme 5a). Smith and co-workers⁷² carefully reexamined the transformation of *d*-**19** and *dl*-**19** by *Curvularia lunata* and confirmed that the 10 β -hydroxy derivative is the major product (*d*-**23**: 55% and 50%), with diminished amounts of the 11 β -hydroxy (*d*-**27**: 17% and *dl*-**27**: 6%), 14 α -hydroxy (*d*-**30**: 13% and 7.3%), and 10 β ,11 β -dihydroxy products (*d*-**28**: 0.8% and *dl*-**28**: 10%) being produced (Schemes 5b and 5c). In addition to these products, they isolated 6 β -hydroxy product (*d*-**22**: 4% and *dl*-**22**: 4.8%), not reported by Szpilfogel.⁷¹ Garrett and co-workers⁷³ found that incubation of 19-nortestosterone with *Aspergillus tamarri* for 72 h gave five transformation products; the major reaction pathway produced 19-nortestolacetone (**33**: 70%). The consecutive steps to the D-ring δ -lactone involve the initial interaction of the steroid substrates with 17-ketodehydrogenase followed by the action of a lactonase enzyme system. An 11 β -hydroxylase enzyme system was also found to be operative on the 19-nortestosterone substrate since 11 β -hydroxy-19-nortestosterone (**27**: 1.8%), 19-norandrost-4-en-



Reagents and Results: a) *Curvularia lunata*, **27**: 3.2%; **23**: 36%; **28**: 0.7%; **30**: 2.3%;
 b) *Curvularia lunata*, **27**: 17%; **22**: 4%; **23**: 55%; **28**: 0.8%; **30**: 13%;
 c) *Curvularia lunata*, *dl*-**27**: 6%; *dl*-**22**: 4.8%; *d*-**23**: 50%; *dl*-**28**: 10%; *l*-**29**: 1.4%; *d*-**30**: 7.3%; *l*-**31**: 4.8%.

Scheme 5

11 β -ol-3,17-dione (**32**: 2.3%), and 11 β -hydroxy-19-nortestololactone (**34**: 3.4%) were obtained (Scheme 6).



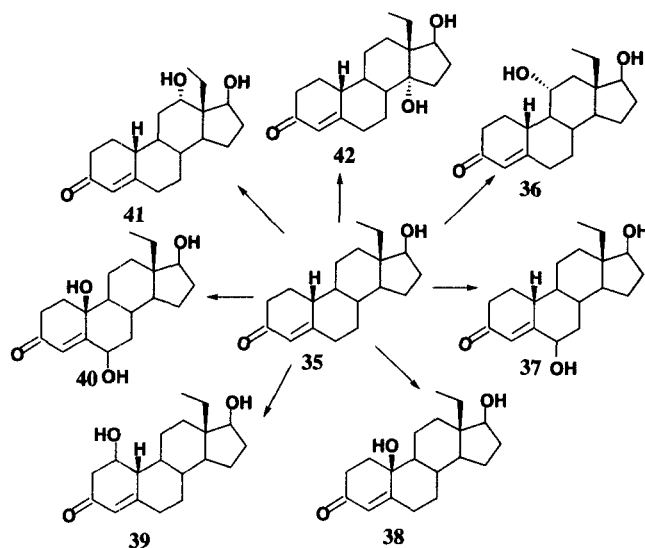
Reagents and Results: *Aspergillus tamarri*, **26**: 3.2%; **32**: 2.3%; **33**: 70%; **34**: 3.4%; **27**: 1.8%.

Scheme 6

c) Fermentation of 13 β -Ethyl-17 β -hydroxygon-4-en-3-one

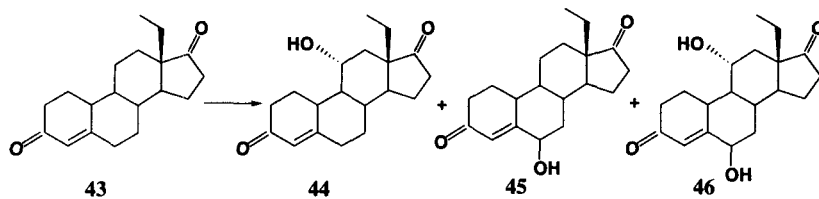
11 α -OH-derivative

Aspergillus ochraceus converted 13 β -ethyl-17 β -hydroxygon-4-en-3-one into a major product (**39**: 52%), together with a series of minor polar components (Scheme 7a).⁷⁰ A trace of 13 β -ethyl-11 α ,17 β -dihydroxygon-4-en-3-one was isolated (**36**: 0.12%). After racemic 13-ethyl-17 β -hydroxygon-4-en-3-one was transformed by *Curvularia lunata*, the gas chromatographic pattern of transformation products showed that two major products (*dl*-**38**: 30% and *l*-**41**: 38.5%) were produced (Scheme 7b);⁷² none of the 11-hydroxy derivative was detected. In 1983, Ulrich and co-workers⁷⁴ used *Aspergillus ochraceus* on 13 β -ethyl-4-ene-3,17-dione **43** and successfully isolated the 11 α -derivative (**44**: 40%). In later work,^{75, 76} the process was repeated and the 6 β - (**45**: 7%) and 11 α ,6 β -dihydroxy (**46**: 13%) derivatives were isolated as well (Scheme 8).



Reagents and Results: a) *Aspergillus ochraceus*, 36: 0.12%; 37: 3.9%; 38: 2.9%; 39: 52%;
 b) *Curvularia lunata*, dl-37: 6.5%; dl-38: 30%; d-40: 1.7%; l-41: 38.5%; d-42: 14%.

Scheme 7

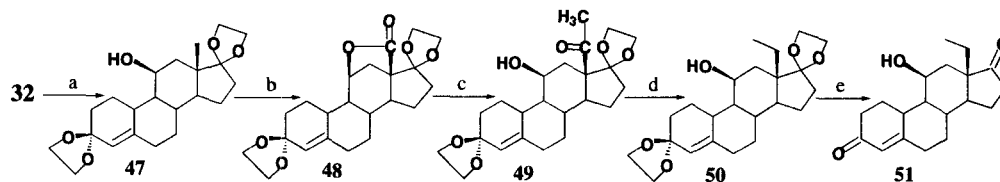


Reagents and Results: *Aspergillus ochraceus*, 44: 42%; 45: 7%; 46: 13%.

Scheme 8

11 β -OH-derivative

A partial synthesis of the 13 β -ethyl-11 β -hydroxygon-4-ene-3,17-dione **51** was reported by Zeelen and co-workers in 1988.⁷⁷ The key step in the synthesis is the intramolecular hypiodite reaction which allows the functionalization of the angular 18-methyl group. Conversion **32** to **51**, required a total of four steps and the overall yield was 33.2% (Scheme 9).

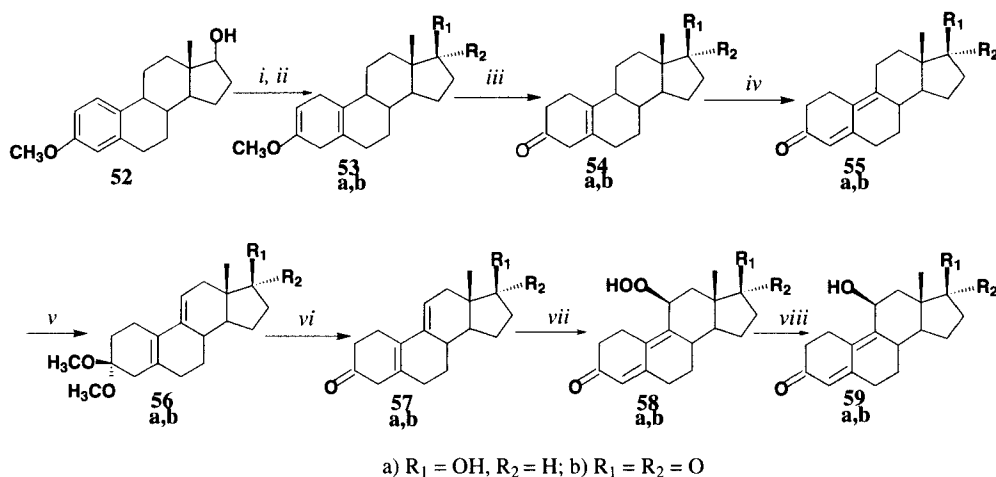


Reagents and Results: a) HC(OEt)₃, (CH₂OH)₂, *p*-TsOH; b) Pb(OAc)₄, I₂, azobisisobutyronitrile, 40%;
 c) CH₃MgBr, ether, quantitative yield; d) Wolff-Kishner reduction 83%; e) *p*-TsOH, acetone, quantitative yield

Scheme 9

2. Oxygen Peroxidation

The oxygen peroxidation of 5(10),9(11)-dien-3-one steroids under the basic condition was first reported by Joly and co-workers.⁷⁸ The intermediate peroxides were unstable and were immediately reduced to hydroxides. Later investigation demonstrated that this is an effective method of introduction 11 β -OH into steroids.⁷⁹⁻⁸² The process starting from 3-methoxy-estra-1,3,5(10)-trien-17-ol (**52**) included eight steps (Scheme 10). Birch reduction of compound **52** gave the 3-methoxy-2,5(10)-diene steroid **53a**,⁸³⁻⁸⁶ subsequent Oppenauer oxidation yielded **53b**. Selective hydrolysis of



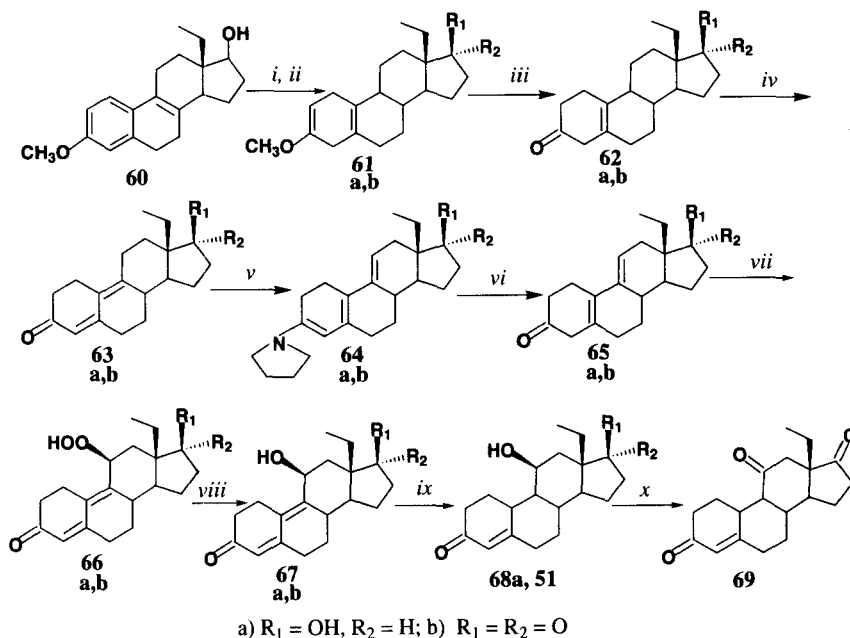
Reagents and Results: i) Li, liq NH₃, ethanol, THF, **53a**: 90%.
 ii) (*i*-PrO)₃Al, cyclohexanone, toluene, **53b**: 87%. iii) Oxalic acid, acetone, **54a**: 83%, **54b**: 85%.
 iv) Pyridinium hydrobromide perbromide (PBP), pyridine, **55a**: 89%, **55b**: 49%.
 v) CH₃COCl, MeOH, **56a**: 95%, **56b**: 97.5%. vi) 8% H₂SO₄, acetone, **57a**: 98%, **57b**: 93%.
 vii) O₂, MeOH (1% Et₃N), **58a**: 100%, **58b**: 92%.
 viii) NaI, HOAc-dioxane, **59a**: 45%; (MeO)₃P, MeOH, **59b**: 82%.

Scheme 10

3-methoxy-2,5(10)-diene steroids (step iii) could be accomplished by using acid catalysts, such as aliphatic acids (e. g. acetic (86%)⁸⁵ and propanedioic acid (100%)⁸⁷) or mineral acids (e. g. perchloric⁸⁸ and sulfuric acid⁸⁵). The most common catalyst was oxalic acid.^{83, 84, 87a, 89} Jones' oxidation of **54a** gave **54b** in a 41% yield.⁹⁰ The 4,9-dien-3-ones (**55**) was formed in one step *via* bromination and dehydrobromination using pyridinium bromide perbromide (PBP) in pyridine.^{89, 91-93} The oxidation of **55** to **55b** could be accomplished with chromic acid;⁹² the same method converted estr-4-ene-3,17-dione-19-carboxy acid to **55b**.^{94, 95} The conversion of 4,9-dien-3-ones into 5(10),9(11)-isomers could be accomplished *via* selective hydrolysis of an enol ester (73%)⁸¹ or enamine⁹⁶, or by tetrachlorosilane (SiCl₄)-catalyzed isomerism.⁹⁷ Enamination with pyrrolidine followed by formic acid-catalysed hydrolysis was used on **55b**,⁹⁵ 17 α -ethynyl-17 β -hydroxy-19-norandrost-4,9(10)-dien-3-one⁹⁸ or similar compounds.⁹⁹ Ketalization with methanol followed by 8% H₂SO₄-catalysed hydrolysis has been used commonly.^{80, 93, 100, 101} Ketalization on a similar compound to **55a** with 2,2-

dimethylpropane-1,3-diol gave 74% yield.¹⁰² The 5(10),9(11)-dien-3-ones were unstable and peroxidised on exposure to oxygen^{78-82, 95, 96, 103, 104} The resulting 11 β -hydroperoxide could be reduced by (MeO)₃P,^{78, 79, 104} NaI^{80, 81, 95} or Na₂SO₃ (86%).¹⁰⁵ in methanolic acetic acid to give **59**.

During the synthesis of desogestrel and its derivatives, Liu and co-workers¹⁰⁶ successfully applied the same strategy to synthesize 13 β -ethyl-gon-4-ene-3,11,17-trione **69** with the total yield of 44% starting from 13 β -ethyl-17 β -hydroxy-3-methoxy-gona-2,5(10)-diene (*Scheme 11*). Hydrolysis of 3-methoxy-2,5(10)-diene steroids (**61a-b**) *via* oxalic acid in presence of silica gel¹⁰⁷ was developed as a rapid and selective synthetic method to produce the corresponding 5(10)-en-3-ones (**62a-b**) in good yields (87-94%).^{106, 108} Steps v and vi could be accomplished simply and in high yields (94-95%) by enamination with pyrrolidine followed by formic acid-catalyzed hydrolysis. Selective hydrogenation of 17-substituted 13 β -ethyl-11 β -hydroxy-gon-4,9-dien-3-ones (**67a-b**) with a



Reagents and Results: (i) Li, liq NH₃, ethanol, THF, **61a**: 63.2%.

(ii) (*i*-PrO)₃Al, cyclohexanone, toluene, **61b**: 85%.

(iii) Oxalic acid on silica gel, CH₂Cl₂, **62a**: 94%, **62b**: 87%.

(iv) Pyridinium hydrobromide perbromide (PBP), pyridine, **63a**: 92%, **63b**: 57%.

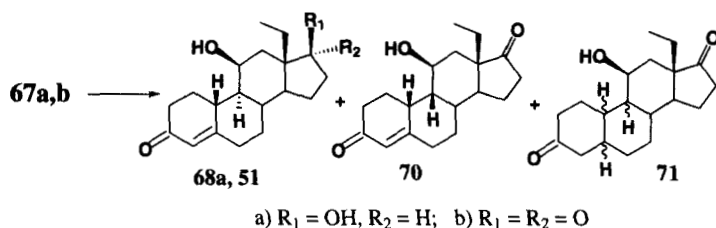
(v) Pyrrolidine, MeOH, **64a**: 94%, **64b**: 95%. (vi) 86% HCOOH. (vii) O₂, MeOH (1% Et₃N).

(viii) NaI, HOAc-dioxane, over-all yield of vi, vii, viii **67a**: 70%, **67b**: 46%

(ix) H₂, 5% Pd/SrCO₃, pyridine, **68a**: 81%, **51**: 45%. (x) Jones' reagent, **69**: 95%.

Scheme 11

Pd(0)/SrCO₃-pyridine medium gave the corresponding 11 β -hydroxy-gon-4-en-3-ones (**68a, 51**) (*Scheme 11* and *12*).^{106, 109} Complete assignments of ¹H and ¹³C NMR spectra of the main products (**68a, 51**) were made by one and two dimensional NMR techniques, such as J-MOD or ATP, ¹H-¹H COSY, HETCOR and COLOC.¹⁰⁹

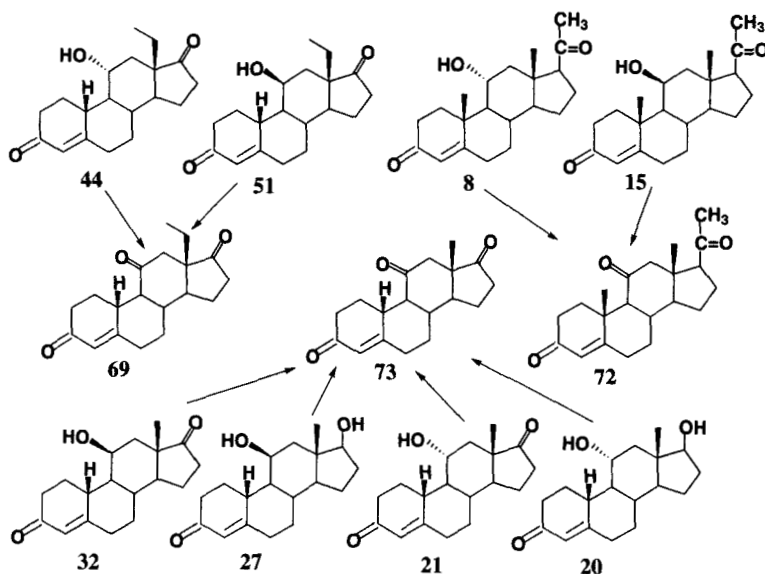


Reagents and Results: H_2 , 5% Pd/SrCO₃, pyridine, **68a**: 81%; **51**: 45%; **70**: 4%; **71**: 4%.

Scheme 12

3. Oxidation of 11-OH Steroids with 4-En-3-ones

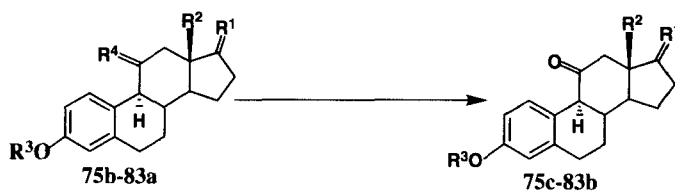
Jones' oxidation of 11-OH-steroids (**8**^{46, 47, 110}, **15**¹¹¹, **20**^{70, 71}, **21**^{71, 11}, **32**⁷⁰, **27**⁷³, **44**^{75, 76}, **51**⁷⁷) yielded the corresponding 11-oxo-steroids (Scheme 13). Jones' reagent can effectively oxidize both



Reagents and Results: Jones' reagents (8N chromic acid), acetone, 0-5°, 15 min. **8**: 67%; **15**: 70%; **20**: 57%; **21**: 82%; **27**: 63%; **32**: 74%; **44**: 83%; **51**: 95%.

Scheme 13

11 α -OH and 11 β -OH. In the meantime, the 17-OH was oxidized to the 17-one. Thus, compounds **8** and **15** gave the same compound **72**; compounds **20**, **21**, **27** and **32** gave the same compound **73**; compounds **44** and **51** gave the same compound **69**. Oxidation of 11 α -hydroxyprogesterone with Na₂Cr₂O₇ gave a 61% yield of 4-pregnene-3,11,20-trione (Scheme 14a).¹¹² 11 α -Hydroxyprogesterone was oxidized to 11-oxoprogesterone (60%) by treatment with *N,N*-diethylamino-1-propyne and DMSO in presence of phosphoric acid (Scheme 14b).¹¹³ In 1965, Moffatt and Pfitzner¹¹⁴ reported that the oxidation of many different types of hydroxy groups, particularly in the steroid area, could be carried out by reaction with dimethyl sulfoxide (DMSO) and dicyclohexylcarbodiimide (DCC) in the

Table 4. Oxidation of Estra-1,3,5(10)-trien-11-ol Derivatives

Compd.	substituents				Reagents and Conditions	Prod. yield (%)	ref.	
	R ¹	R ²	R ³	R ⁴				
75b		CH ₃	CH ₃	α-OH, β-H	1. DCC, DMSO, Cl ₂ CHCOOH; 2. PCC/CH ₂ Cl ₂	75c	100	120
						75c	28	121
77b	β-OCH ₂ Ph	C ₂ H ₅	CH ₃	α-OH, β-H	1. PCC/CH ₂ Cl ₂ ; 2. DCC/DMSO	77c	64	129
						77c		131
80b	β-OCH ₂ Ph	CH ₃	PhCH ₂	α-OH, β-H	1. PDC/DMF; 2. PCC/CH ₂ Cl ₂	80c	16	136
						80c	57	4
81a	β-OCH ₂ Ph	C ₂ H ₅	PhCH ₂	α-OH, β-H	1. PDC/DMF	81b		136
82a	=O	CH ₃	OH	β-OH, α-H	1. CrO ₃ /H ₂ SO ₄	82b	38	135
83a		CH ₃	PhCH ₂	α-OH, β-H	1. PCC/CH ₂ Cl ₂	83b	64	123

Additional applications of PCC to the oxidation of very similar compounds showed the satisfactory results (**83b**: 64%,¹²³ **80c**: 57%,⁴ **77c**: 64%¹²⁹). Coombs and co-workers¹²⁰ oxidized **75b** with dicyclohexylcarbodiimide and DMSO in presence of dichloroacetic acid to 11-one with 100% yield. The Moffatt oxidation reaction was also used on compound **77b** but there were difficulties in purification of the products because of DCC and DCU residues.^{131, 137}

b) Hydroboration-Chromic Acid Oxidation

The chromic acid oxidation of organoboranes to yield ketones was first reported by Brown and Garg.¹³⁸ Wang and Li used the same method on 13β-ethyl-3-methoxygona-1,3,5(10),9(11)-tetraene-17β-ol **76a** and isolated four products (Scheme 19).^{139, 140} The ¹H NMR data for these compounds are summarized in Table 5. For the compound **76h**, MS and elemental analysis showed the molecular formula of C₂₀H₂₇BO₄. The IR spectrum showed two peaks for hydroxy groups (3496 cm⁻¹ and 3306 cm⁻¹) and one peak for a ketone (1718 cm⁻¹). The ¹³C NMR showed the same carbon number as that of **76a**. With DEPT technique, there were still multiple peaks at 24.60 ppm. According to the mechanism suggested,¹³⁸ **76h** was supposed to be 11-boron derivative. ¹¹B NMR gave a peak at 15.62 ppm (B₂H₆ as standard). After alkaline peroxide oxidation, the product was identified with **76g**. Thus, the structure of **76h** was 13β-ethyl-3-methoxygona-1,3,5(10)-trien-17-one-11α-boron acid. For the compound **76i**, MS gave the molecular weight of 294. ¹H NMR showed five aromatic protons: 7.88 (1H, d, J = 9 Hz, 6-H), 7.64 (1H, d, J = 8.5 Hz, 1-H), 7.18 (1H, d, J = 9 Hz, 7-H), 7.15 (2H, m, 2,

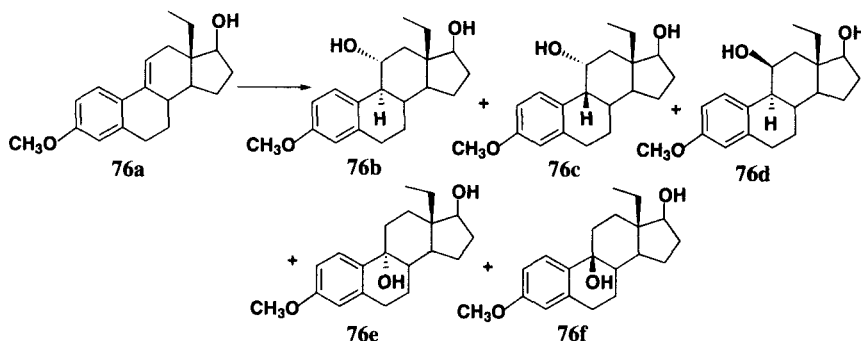
II. SYNTHETIC APPROACHES TO 11-OXO STEROIDS WITH AROMATIC A-RING

1. Hydroboration/Oxidation

a) Hydroboration-Alkaline Peroxide Oxidation

Hydroboration

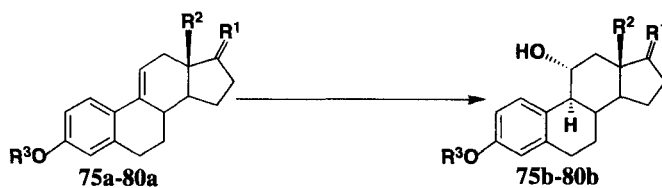
After treating 17,17-ethylenedioxy-3-methoxyestra-1,3,5(10),9(11)-tetraene **75a** (Table 1) with diborane in the usual way¹¹⁹, Coombs and co-workers isolated a product formulated as the 11 α -hydroxy-derivative **75b**.¹²⁰ The process was repeated successfully later and **75b** was obtained in 61% yield.¹²¹ Using the same strategy on 13 β -ethyl-3-methoxygona-1,3,5(10),9(11)-tetraene-17 β -ol **76a**, Smith isolated two products (**76b** and **76e**) (Scheme 16).¹²² In 1985, Li and co-workers¹²³ isolated a



Reagents and Results: BH_3/THF , $\text{H}_2\text{O}_2\text{-NaOH}$, **76b**: 56%; **76c**: 10%; **76d**: 2%; **76e**: 0.4%; **76f**: 3%.

Scheme 16

Table 1. Hydroboration-Alkaline Peroxide Oxidation of Estra-1,3,5(10),9(11)-tetraene Derivatives



Compd	substituents			Reagents and Conditions	Prod. yield (%) ref		
	R ¹	R ²	R ³				
75a		CH ₃	CH ₃	1. BH_3/THF 2. $\text{H}_2\text{O}_2\text{-NaOH}$	75b	35, 61	121
76a	$\beta\text{-OH}$, $\alpha\text{-H}$	C ₂ H ₅	CH ₃	1. BH_3/THF 2. $\text{H}_2\text{O}_2\text{-NaOH}$; 1. $\text{NaBH}_4/\text{BF}_3 \text{ Et}_2\text{O}$ 2. $\text{H}_2\text{O}_2\text{-NaOH}$	76b	56	122
77a	$\beta\text{-OCH}_2\text{Ph}$	C ₂ H ₅	CH ₃	1. BH_3/THF 2. $\text{H}_2\text{O}_2\text{-NaOH}$	77b	49	129
78a	=O	C ₂ H ₅	CH ₃	1. BH_3/THF 2. $\text{H}_2\text{O}_2\text{-NaOH}$	78b	a	131
79a	=O	CH ₃	CH ₃	1. $\text{NaBH}_4\text{-AlCl}_3$ 2. $\text{H}_2\text{O}_2\text{-NaOH}$	79b	a	133
80a	$\beta\text{-OCH}_2\text{Ph}$	CH ₃	PhCH ₂	1. $\text{LiAlH}_4/\text{BF}_3 \text{ Et}_2\text{O}$ 2. $\text{H}_2\text{O}_2\text{-NaOH}$; 1. $\text{LiBH}_4/\text{catecholborane}$ 2. $\text{H}_2\text{O}_2\text{-NaOH}$	80b	a	134
					80b	72	4

a) Yields were not reported.

third isomer **76d** in the same reaction mixture. Three years later, Zhou and Li¹²⁴ carefully reexamined the same reaction mixture and isolated all five isomers by column chromatography. The ¹H NMR data and the ¹³C NMR data of the five isomers (**76b-76f**) are summarized in Tables 2 and 3, respectively.

Table 2. Partial ¹H NMR Spectra of **76b**, **76c**, **76d**, **76e** and **76f**

Cmpd	3-OCH ₃	17 α -H	C ₁₁ -H	C ₄ -H	C ₂ -H	C ₁ -H
76b	3.75(s)	3.73(s)	4.04(td)	6.60(d)	6.75(dd)	7.82(d)
76c	3.79(s)		4.52(dt)	6.68-7.02(m)	6.68-7.02(m)	7.11(d)
76d	3.73(s)		4.72(broad)	6.62(d)	6.75(dd)	7.16(d)
76e	3.80(s)			6.67, 6.68(m)	6.67, 6.68(m)	7.16(d)
76f	3.79(s)			6.65-6.77(m)	6.65-6.77(m)	7.19(d)

Table 3. ¹³C NMR Spectra of **76b**, **76c**, **76d**, **76e** and **76f**

Assignment	76b	76c	76d	76e	76f
C-1	127.1	129.4	125.9	126.5	125.5
C-2	110.9	112.6	112.2	111.6	111.6
C-3	157.7	157.7	157.7	157.4	157.8
C-4	113.7	113.1	114.7	113.5	113.7
C-5	139.0	135.1	139.9	137.7	136.4
C-6	28.6	34.2	28.9	30.3	29.7
C-7	27.0	30.2	26.7	27.4	26.9
C-8	37.0	43.7	33.0	39.4	39.6
C-9	(50.9)	49.0	(51.8)	85.7	72.2
C-10	132.5	133.1	127.7	132.0	132.7
C-11	70.4	68.9	67.4	25.3	25.9
C-12	43.5	41.5	44.1	31.7	41.3
C-13	45.4	43.8	37.7	51.9	47.3
C-14	(50.2)	44.7	(50.1)	43.8	45.3
C-15	22.5	24.9	22.3	22.8	24.7
C-16	30.8	30.7	30.8	30.9	31.0
C-17	83.1	84.4	84.8	79.7	83.3
C-18	18.4	19.1	20.3	16.7	17.6
C ₁₈ -CH ₃	9.7	10.0	11.1	8.4	9.3
C ₃ -OCH ₃	55.1	55.2	55.1	55.1	55.2

Compound **76b**, **76c** and **76d** were identified by their melting points and ¹H NMR spectra data.^{122, 125} When 11-OH is at the α -position (**76b**), the 11 β -H is deshielded to 4.04ppm. Among the 11 β -H, 9 α -H, 12 α -H and 12 β -H, the first three hydrogens are axial and the last one is equatorial.

Thus, both $J_{9\alpha, 11\beta}$ and $J_{11\beta, 12\alpha}$ are 10 Hz which is larger than $J_{11\beta, 12\beta}$ (5 Hz). The 11 β -H shows a triple-doublet (Fig. 2). When the 11-OH is α oriented and the 9-H is β oriented (**76c**), the 11 β -H is shifted to 4.52 ppm. Among the 11 β -H, 12 α -H, 9 β -H and 12 β -H, the first two hydrogens are axial and the last

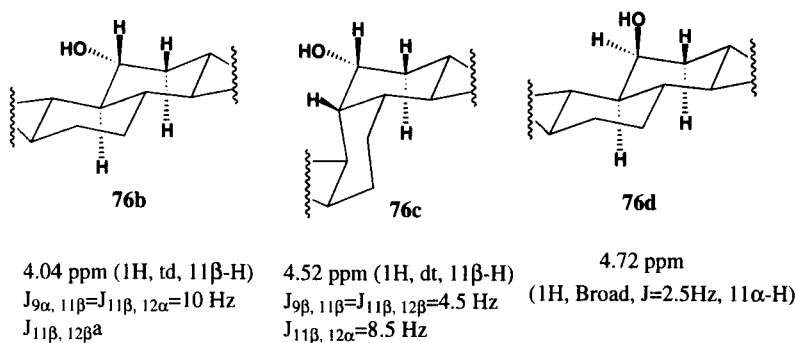
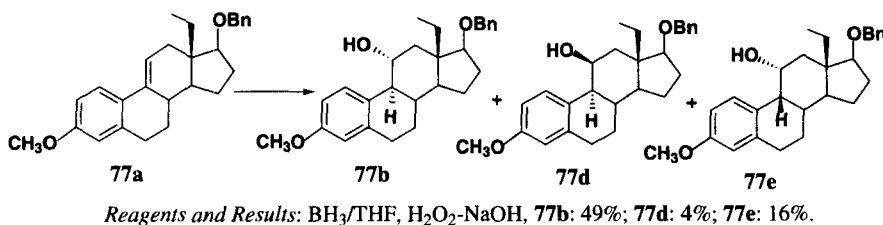


Fig. 2 ^1H NMR coupling configuration determination of **76b**, **76c** and **76d**.

two are equatorial. Thus, $J_{9\beta, 11\beta}$ and $J_{11\beta, 12\beta}$ are 4.5 Hz which is smaller than $J_{11\beta, 12\alpha}$ (8.5 Hz). The 11 β -H shows a double-triplet. When 11-OH is at β -position (**76d**), the 11 α -H deshields to 4.72 ppm and shows a broad peak with $J = 2.5$ Hz. This is due to 60° angle between 11 α -H and each of 9 α -H, 12 α -H and 12 β -H. For compound **76e**, ^1H NMR spectrum did not show any peaks between 4 ppm and 5 ppm, and it easily dehydrated in $\text{HCl-CH}_3\text{OH}$ at room temperature to give $\Delta^{9(11)}$. There was positive Cotton effect at 228 nm. All these data demonstrated that **76e** had a 9-OH at the α -position. In the ^{13}C NMR, the two tertiary carbon peaks appeared at C-17 (79.7 ppm) and C-9 (85.7 ppm). For compound **76f**, the elemental analysis and MS spectra were consistent with the formula $\text{C}_{20}\text{H}_{28}\text{O}_3$. In ^1H NMR, there were no peaks between 4 ppm and 5 ppm which indicated there was no other CH(OH) grouping in the molecule except 17 β -OH. Refluxing **76f** in $\text{HCl-CH}_3\text{OH}$ produced a $\Delta^{9(11)}$ double bond and showed that a second hydroxy group was connected to C-9. In ^{13}C NMR, the two tertiary carbon peaks assigned to C-17 (83.3 ppm) and C-9 (72.2 ppm). The negative Cotton effect at 228 nm determined the 9-OH to be in the β -configuration.^{126, 127} The introduction of the 9-OH is clearly activated by the A-ring aromatic system. The assignment of C, CH, CH_2 and CH_3 was accomplished by INEPT technique based reference compounds and substituent effects.¹²⁸ For compound **76d**, there is γ relationship between C-11 β -OH and C-8, C-13 with cross conformation. Compared with **76b**, $\delta_{\text{C-8}}$ and $\delta_{\text{C-13}}$ were more shielded (4.0 and 7.7 ppm respectively). The three tertiary carbons (C-8, C-9 and C-14) in **76c** were assigned by conformation. Because the C-14H approached the C-6H, $\delta_{\text{C-14}}$ was deshielded to higher field (44.7 ppm). The relationship between C-8H and C-6H changed from axial-axial to axial-equatorial causing $\delta_{\text{C-8}}$ to be deshielded (43.7 ppm). Because 9 α -OH of compound **76e** showed γ effect, $\delta_{\text{C-12}}$ and $\delta_{\text{C-14}}$ were more shielded (9.6 and 1.5 ppm respectively) compared with compound **76f**. In the synthesis of 3-oxo desogestrel, Gao and co-workers applied the same method on the similar compound **77a** and isolated three products with 11 α -hydroxy derivative (**77b**: 49%) as the major product (Scheme 17).^{129, 130} This method has also been used on the related compound with a 17-one

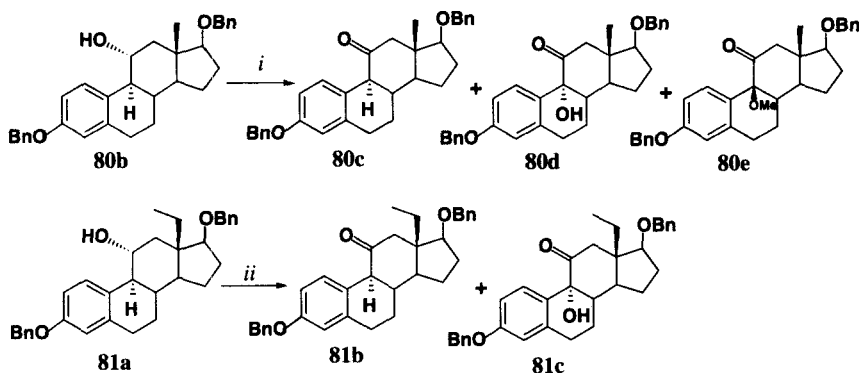
group **78a**.¹³¹ To avoid the complexity in the preparation of diborane solution in THF, other more convenient methods such as $\text{NaBH}_4\text{-BF}_3\cdot\text{Et}_2\text{O}$,¹³² $\text{NaBH}_4\text{-AlCl}_3$,¹³³ $\text{LiAlH}_4\text{-BF}_3\cdot\text{Et}_2\text{O}$,¹³⁴ $\text{LiBH}_4\text{-catecholborane}$ ⁴ gave better yields (Table 1).



Scheme 17

Oxidation

The proper oxidant should be selected to oxidize estra-1,3,5(10)-trien-11-ol derivatives. Because C-9 is adjacent to C-11 and is also benzylic, 9 α -OH-11-ketone may be obtained along with the anticipated 11-ketone. Oxidation 11 β -hydroxyestrone **82a** with Kiliani's reagent gave a 38% of 11-ketoestrone **82b** (Table 4).¹³⁵ In 1985, Hanson and co-workers¹³⁶ reported that the oxidation of 3,17 β -dibenzoyloxy-11 α -hydroxyestra-1,3,5(10)-triene **80b** with pyridinium dichromate (PDC) yielded the corresponding 9 α -hydroxy-11-ketone **80d** as well as the anticipated 11-ketone **80c**. The 9 β -methyl ether **80e**, obtained as a result of a methanolic work-up and shown to arise by an acid-catalyzed methanolysis of the 9 α -alcohol, had its stereochemistry confirmed by X-ray crystallographic analysis (*Scheme 18i*). When the alcohol **81a** containing a 13-ethyl substituent was subjected

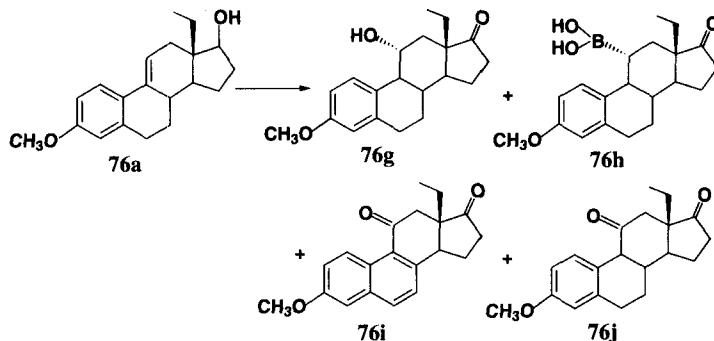


Reagents and Results: i) DCC, DMF, MeOH, **80c**: 16%; **80d**: 1.2%; **80e**: 41%; ii) DCC, DMF, **81b** and **81c**.

Scheme 18

to the same oxidation sequence but without treatment with methanol, the corresponding ketone **81b** and ketol **81c** were obtained (*Scheme 18ii*). Oxidation of 17,17-ethylenedioxy-3-methoxyestra-1,3,5(10)-trien-11 α -ol **75b** with pyridinium chlorochromate (PCC) afforded the non-crystalline 11-ketone **75c** (Table 4).¹²¹

4-H). ^{13}C NMR showed five aromatic carbons between 100 and 200 ppm. Thus, the structure of **76i** was 13 β -ethyl-3-methoxygona-1,3,5(10),6,8-pentaen-17-one. The anticipated product **76j** showed two



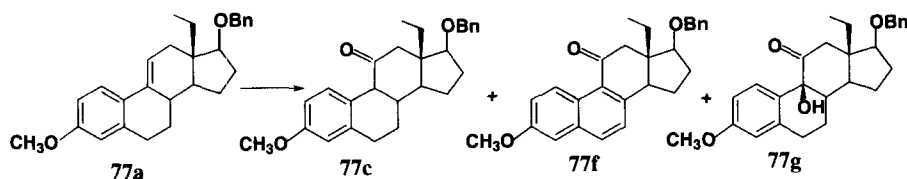
Reagents and results: a) BH_3/THF , rt., 48 h; b) H_2CrO_4 , rt., 5h. **76g**: 9%, **76h**: 32%, **76i**: 13%, **76j**: 28%.

Scheme 19

Table 5. Partial ^1H NMR Spectra of **76g**, **76h**, **76i** and **76j**

Cpd.	3-OCH ₃	C ₁₁ -H	C ₄ -H	C ₂ -H	C ₁ -H	C ₆ -H	C ₇ -H
76g	3.78(s)	4.02(td)	6.64(m)	6.64(m)	7.99(d)	—	—
76h	3.70(s)	6.62(m)	6.62(m)	7.18(d)	—	—	—
76i	3.94(s)	—	7.15(m)	7.15(m)	7.64(d)	7.88(d)	7.18(d)
76j	3.78(s)	—	6.60-6.84(m)	7.27(d)	—	—	—

carbonyl absorptions at 1736 cm^{-1} and 1714 cm^{-1} . The C₉-H showed a doublet ($J_{8,9} = 9.8$ Hz) at 3.52 ppm. The 9.8 Hz coupling constant confirmed the 9 α -configuration.¹⁴¹ In this reaction, a TLC analysis showed that **76h** and **76i** were formed at the very beginning of the reaction. With the different molar ratios of oxidant and reaction temperatures, compound **76h** was still the major product. Using the same method on the compound **77a**, the yield of anticipated product **77c** was improved to 47% (Scheme 20).^{139, 142} For the compound **77g**, the result of MS and elemental analysis indicated a molecular formula $\text{C}_{27}\text{H}_{32}\text{O}_4$. ^1H NMR did not show the peaks typical of 9H-11-one structure between 3.40 and 3.60 ppm. The three peaks in ^{13}C NMR between 70 and 100 ppm demonstrated 9-OH structure while the other two peaks belonged to 17-OCH₂- and C-17. The earlier study showed that 9 β -hydroxy



Reagents and Results: a) BH_3/THF , rt., 48h; b) H_2CrO_4 , rt., 2h. **77c**: 47%, **77f**: 13%, **77g**: 17%.

Scheme 20

group resulted in *cis*-B/C rings and formed hydrogen bond with 11-one (Fig. 3 A).¹⁴³ This resulted in C₁-H being shielded to the higher field. The chemical shift of C₁-H (6.60-6.67 ppm, 3H, m, 1, 2, 4-H) in ¹H NMR and IR frequency of 9-OH group (3432 cm⁻¹) confirmed the 9β-OH configuration.

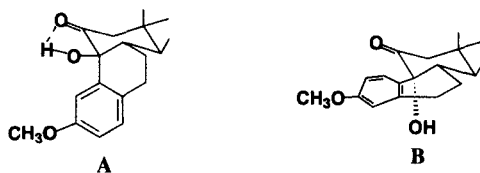


Fig. 3

2. Epoxidation

The epoxidation of 9(11)-dehydroestrone acetate **84a** with perbenzoic acid in chloroform afforded a mixture of an epoxide.¹⁴⁴ The epoxide obtained as the major product (72%) was identified to have the 9α,11α-epoxide configuration, whereas the minor product (12%) has the 9β,11β-epoxide configuration (Table 6). Liang and co-workers¹⁴⁵ used *m*-chloroperbenzoic acid to epoxidise **85a** and got an epoxide mixture with 80% yield. Upon epoxidation of **75a** by the two-phase dichloromethane/aqueous potassium carbonate procedure of Anderson and Veysoglu,¹⁴⁶ Collin and co-workers^{121, 147} obtained a 70% yield of crystalline but extremely acid-sensitive 9α,11α-epoxide **75d**. Suitable precautions were necessary for its recrystallization and spectroscopic examination.¹²¹



Table 6. Epoxidation of Estra-1,3,5(10),9(11)-tetraene Derivatives

Compd	substituents		Reagents and Conditions	yield (%)			ref
	R ¹	R ²		9a,11a	9b,11b	9z,11z	
84a	=O	AcO	1. PhCO ₂ OH, CHCl ₃ , 0°, 24 h	84b : 72	84c : 12		144
85a	β-OAc, α-H	AcO	1. <i>m</i> -ClPhCO ₂ OH, CHCl ₃ , 0°, 2 h			85b : 80	145
75a		OCH ₃	1. <i>m</i> -ClPhCO ₂ OH, NaHCO ₃ , CHCl ₃	75d : 70-72			121

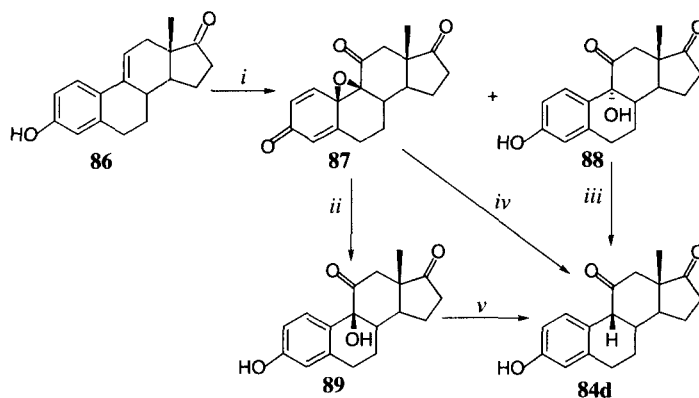
Treatment of 9α,11α-epoxides with dry hydrogen chloride in methanol¹⁴¹ or 5% KOH in methanol¹⁴⁵ afforded the corresponding 3-hydroxy-9β-estra-1,3,5(10)-trien-11-one derivatives (Table 7) *via* a pinacol-pinacolone-like mechanism. A solution of **75d** in benzene containing a trace of anhy-

drous lithium perchlorate was heated under reflux for 3 hours to give a nearly quantitative yield of the non-crystalline 9 β -ketone **75e**.^{121, 147}

Table 7. Conversion of 9a,10a-Epoxides to 11-Ones

Compd	substituents		Reagents and Conditions	substituents		Prod. yield (%)	ref
	R ¹	R ²		R ¹	R ²		
84b	=O	AcO	1. HCl, MeOH, -5°, 1.5 h	=O	HO	84d	80 141
85b	β -OAc, α -H	AcO	1. KOH, MeOH, reflux, 30 min.	β -OH, α -H	HO	85c	66 145
75d		OCH ₃	1. LiClO ₄ , benzene, reflux, 3 h		OCH ₃	75e	98-100 121

Hasegawa and Tsuda's further work demonstrated that the treatment of $\Delta^{9(11)}$ -estrone **86** with three molar equivalents of perbenzoic acid in chloroform afforded an epoxydienone **87** in 40-50% yields and a phenolic compound **88** in 3-10% yields (Scheme 21).¹⁴³ When **87** was treated with zinc in



Reagents and Results: (i) PhCO₂OH, CHCl₃, 0°, 24 h, **87**: 44%, **88**: 4.7%; (ii) H₂, 5% Pd-C, **89**: 50%; (iii) Zn/HOAc, reflux, 4 h, **84d**: 21%; (iv) Zn/HOAc, reflux, 2 h, **84d**: 15%; (v) Zn/HOAc, reflux, 19 h, **84d**: 22%.

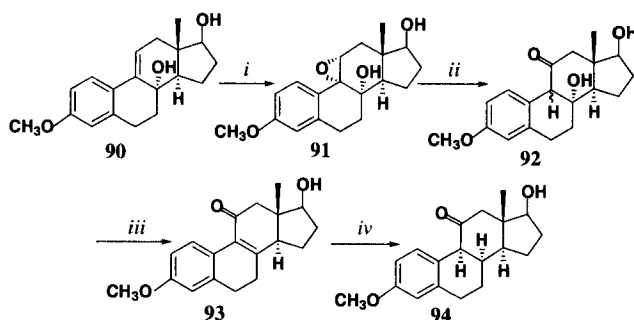
Scheme 21

acetic acid, 3-hydroxy-9 β -estra-1,3,5(10)-triene-11,17-dione **84d** was obtained. On hydrogenation with 5% palladium-charcoal **87** readily consumed one molar equivalent of hydrogen to afford in high yield a phenolic compound determined to be 3,9-dihydroxyestra-1,3,5(10)-triene-11,17-dione (**89**). Elemental analysis indicated that **88** was isomeric with **89**.

The hydroxy group of **88** was resistant to oxidation by chromic anhydride in pyridine and thus might be tertiary. Reduction of **88** with zinc in acetic acid resulted in hydrogenolysis to yield **84d**

as did **87**. The configurations of the hydroxy groups of **88** and **89** was assigned by measurement of intramolecular hydrogen bonding in the infrared spectra. The hydroxy and carbonyl absorption bands in dilute carbon tetrachloride solution ($c = 0.0004 \text{ M}$, $l = 50 \text{ mm}$) of **88** and **89** were observed at 3609, 1730 and 3843, 1713 cm^{-1} , respectively. The later frequencies indicated the presence of intramolecular hydrogen bonding between C_9 -hydroxyl and C_{11} -carbonyl group, while the former did not. Inspection of Dreiding Model as shown in Fig. 3 indicates that the 9β -hydroxy isomer (A) should exhibit hydrogen bonding, while the 9α -hydroxy isomer (B) cannot. Thus the α - and β -configuration were assigned to the hydroxy group of **88** and **89** respectively.

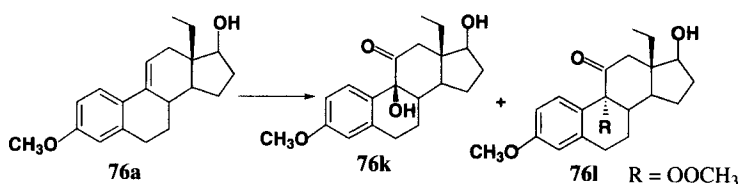
When *m*-chloroperbenzoic acid was used on the diol **90** in benzene-hexane, the epoxydiol **91** was obtained (Scheme 22).¹⁴⁸ Methanolic HCl at or below room temperature transformed **91** to 11-oxo-estratetraene **93**, probably through formation and dehydration of the ketol **92**. The overall yield for the three stage sequence *i-iii* was 52%. Catalytic hydrogenation over palladized charcoal in dimethylformamide converted **93** to an estratriene **94**.



Reagents and Results: (i) *m*-ClPhCO₂OH, benzene-hexane; (ii)-(iii) HCl/MeOH, rt., **90-93**: 52%; (iv) H₂/Pd-C, DMF

Scheme 22

During the synthesis of 3-oxo desogestrel, Wang and Li^{139, 149} attempted a *m*-chloroperbenzoic acid epoxidation on 13 β -ethyl-3-methoxygona-1,3,5(10),9(11)-tetraen-17 β -ol **76a** (Scheme 23).



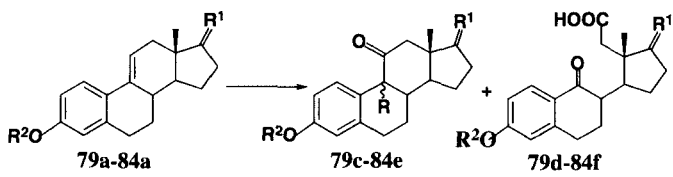
Reagents and Results: (i) *m*-ClPhCO₂OH, CHCl₃, 0°, 2 h; (ii) KOH, MeOH, reflux, 30 min. **76k**: 39%; **76l**: 15%.

Scheme 23

Because of the instability of the $9\alpha,11\alpha$ -epoxide, it was treated with 5% KOH in methanol directly. Column chromatography afforded two compounds. The elemental analysis and MS showed that the compound **76k** had a formula of $\text{C}_{20}\text{H}_{26}\text{O}_4$. The IR spectrum gave two peaks for hydroxy

groups (3458 cm^{-1} and 3440 cm^{-1}) and one ketone peak (1705 cm^{-1}). ^1H NMR spectrum did not give a peak for the $\text{C}_9\text{-H}$ ($\delta=3.40\text{-}3.60\text{ ppm}$). ^{13}C NMR showed a new tertiary-carbon peak at $\delta=85.5\text{ ppm}$. The above analysis showed the presence of a 9-OH structure. The 9β -configuration was identified like **77g**. The elemental analysis and MS demonstrated that compound **76l** had a formula of $\text{C}_{21}\text{H}_{28}\text{O}_5$. Both ^1H NMR ($\delta = 3.39\text{ ppm}$) and ^{13}C NMR ($\delta = 102.4\text{ ppm}$) showed the 9- OOCH_3 . The 9α -configuration was identified by Hasegawa and Tsuda's method.¹⁴³

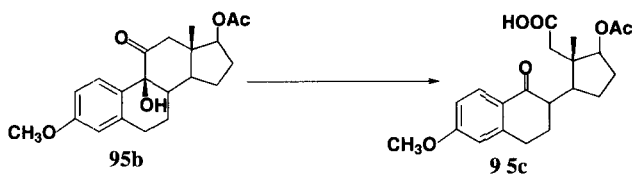
Table 8. Jones' Oxidation of Estra-1,3,5(10),9(11)-tetraene Derivatives



Compd	substituents		Reagents and Conditions	yield (%)		ref
	R ¹	R ²		79c-84e	79d-84f	
79a	=O	CH ₃	1. CrO ₄ -H ₂ SO ₄ , rt., 11 h, R = 9β-OH	79c : 23	79d : 31	153
95a	β-OAc, α-H	CH ₃	1. CrO ₄ -H ₂ SO ₄ , rt., 11 h, R = 9β-OH	95b : 37	95c : 35	153
95a	β-OAc, α-H	CH ₃	1. CrO ₄ -H ₂ SO ₄ , 0°, 5 h, R = 9β-OH	95b : 32	95c : 42	155
96a	H ₂	CH ₃	1. CrO ₄ -H ₂ SO ₄ , rt., 3 h, R = 9β-OH	96b : 11	96c : 4	154
96a	H ₂	CH ₃	1. CrO ₄ -H ₂ SO ₄ , rt., 11 h, R = 9β-OH	96b : 35	96c : 28	153
84a	=O	Ac	1. CrO ₄ -H ₂ SO ₄ , rt., 11 h, R = 9α-OH	84e : 28	84f : 18	153

3. Jones' Oxidation

Jones' oxidation of estra-1,3,5(10),9(11)-tetraene could yield 9β -OH-11-one (**79c-84e**) and 9-oxo-9,11-seco-11-oic acid derivatives (**79d-84f**) (Table 8).¹⁵⁰⁻¹⁵⁵ It was suggested that the 9β -hydroxy-11-oxo-derivatives were formed *via* a 9(11)-dehydro-derivative. Further oxidation of **95b** with sodium metaperiodate produced **95c** nearly quantitatively (Scheme 24).¹⁵⁵ The only 9α -hydroxy-11-oxo derivative was found in the oxidation of 17-oxo-estra-1,3,5(10),9(11)-tetraen-3-yl acetate.¹⁵³ The 9α -H-11-oxo and 9β -H-11-oxo derivatives were reported by Acinou and co-workers.¹⁵⁴



Reagents and Results: NaIO₄, HCl, acetone, reflux, 8 h, **95c**: 98%

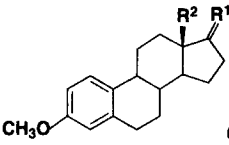
Scheme 24

4. ¹H NMR Studies of Estra-3-methoxy-1,3,5(10)-trien Derivatives

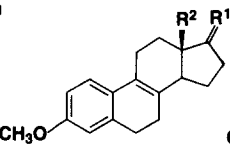
Nuclear magnetic resonance spectroscopy (NMR) is one of the more powerful tools available to the chemist for elucidating the structure of steroids. The steroid skeleton has many aliphatic carbons and protons which show overlapping peaks at high field. However, the special carbons and protons of steroid structures give characteristic NMR signals because of its highly asymmetric structure and distinct chemical environments. In principle, they can be uniquely assigned to give remarkably detailed information on the system.

For the estra-3-methoxy-1,3,5(10)-trien steroids, C₂-H and C₄-H are less deshielded by the aromatic A-ring compared with C₁-H because of back-donation by the 3-methoxy group. Because of the coupling with C₂-H, C₁-H show a doublet (J = 9 Hz). Besides the coupling with C₁-H, C₂-H has a long distance coupling with C₄-H (J = 1.8 Hz) and shows a double-doublet peak. Thus, C₄-H shows a doublet. After the introduction of Δ⁹⁽¹¹⁾, C₁-H is in the plane of A-ring and Δ⁹⁽¹¹⁾. Therefore, it will be deshielded to lower field about 0.45 ppm because the effect from A-ring and Δ⁹⁽¹¹⁾ (Table 9). On the

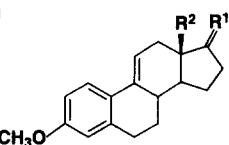
Table 9. ¹H NMR Chemical Shifts of Estra-3-methoxy-1,3,5(10)-triene Derivatives





97-101



102-103

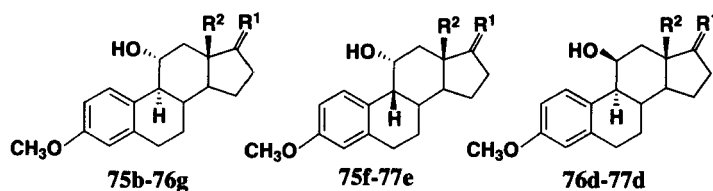


96a-107

Compd.	substituents		Chemical Shifts (ppm)				ref.
	R ¹	R ²	C ₁ -H	C ₂ -H	C ₄ -H	C ₁₁ -H	
97	β-OH, α-H	C ₂ H ₅	7.01	6.53	6.45		131
98	β-OCH ₂ Ph	C ₂ H ₅	7.02	6.63	6.50		139
99	=O	CH ₃	7.12	6.60	6.56		143
100	β-OC(CH ₃) ₃	CH ₃	7.20	6.63	6.63		156
101	β-OC(CH ₃) ₃	C ₂ H ₅	7.18	6.68	6.68		156
102	β-OH, α-H	CH ₃	7.13	6.66	6.66		156
60	β-OH, α-H	C ₂ H ₅	7.11	6.72	6.60		139
103	β-OCH ₂ Ph	C ₂ H ₅	7.16	6.66	6.56		139
96a	=H ₂	CH ₃	7.39	6.70	6.61	6.08	154
75a		CH ₃	7.47	6.48-6.70	6.48-6.70	6.08	121
104	β-OH, α-H	CH ₃	7.53	6.66	6.66	6.11	156
76a	β-OH, α-H	C ₂ H ₅	7.44	6.65	6.54	6.04	139
77a	β-OCH ₂ Ph	C ₂ H ₅	7.45	6.66	6.56	6.04	139
105	β-O ₂  , α-H	C ₂ H ₅	7.45	6.66	6.54	6.04	139
106	β-OC(CH ₃) ₃	CH ₃	7.48	6.58	6.58	6.07	156
107	β-OC(CH ₃) ₃	C ₂ H ₅	7.45	6.63	6.63	6.07	156

other hand, $\Delta^{8(9)}$ cannot become coplanar with A-ring because of the B/C bridge bond. The C_1 -H is only deshielded about 0.1 ppm (Table 9). The introduction of 11α -hydroxy group will deshield C_1 -H by about 0.8 ppm (Table 10). With 9β -H, there will be *cis*-B/C conformation and longer distance

Table 10. ^1H NMR Chemical Shifts of Estra-3-methoxy-1,3,5(10)-trien-11-ol Derivatives



Compd	substituents		Chemical Shifts (ppm)				ref.
	R ¹	R ²	C ₁ -H	C ₂ -H	C ₄ -H	C ₁₁ -H	
75b		CH ₃	7.80	6.56-6.74	6.56-6.74	4.24	121
76b	β -OH, α -H	C ₂ H ₅	7.82	6.75	6.60	4.04	124
77b	β -OCH ₂ Ph	C ₂ H ₅	7.80	6.78	6.72	4.10	129
76g		C ₂ H ₅	7.99	6.65	6.61	4.01	139
75f		CH ₃	7.76	6.68	6.65	4.45	121
76c	β -OH, α -H	C ₂ H ₅	7.11	6.86	6.66	4.52	124
77e	β -OCH ₂ Ph	C ₂ H ₅	7.12	6.78	6.68	4.56	129
76d	β -OH, α -H	C ₂ H ₅	7.16	6.75	6.62	4.72	124
77d	β -OCH ₂ Ph	C ₂ H ₅	6.69	6.62-6.75	6.62-6.75	4.51	129

between C_1 -H and 11α -OH. The C_1 -H are not affected by 11α -OH. From the data, the 11β -hydroxy group do not affect C_1 -H either. The introduction of 11 -one will deshield C_1 -H by about 0.1 ppm (Table 11). On the other hand, C_1 -H will be shielded to the higher field about 0.4 ppm with 9β -configuration and overlap with C_2 -H and C_4 -H.

III. SYNTHETIC APPROACHES TO 11-OXO STEROIDS OF CHOLIC ACID DERIVATIVES

1. Marker-Lawson Method

11-Oxo-deoxycholic Acid Derivatives

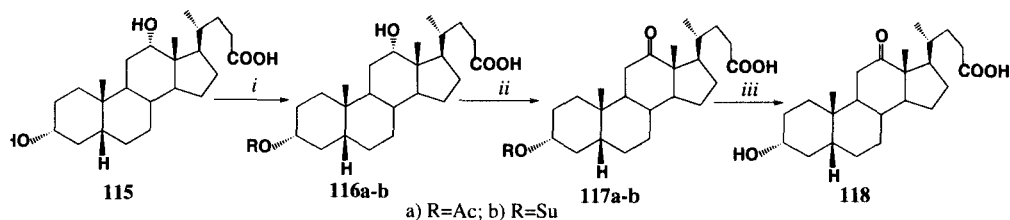
In 1938, Marker and Lawson¹⁵⁷ brominated 3α -acetoxy-12-ketocholanate and then hydrolyzed the resulting 11α -bromoketone to a ketol acid which contained an oxygen function at C-11. This has become known as the Marker-Lawson acid, which is a useful precursor 11-oxygenated steroid.

Desoxycholic acid **115** was treated with acetic anhydride^{157, 158} or succinic anhydride¹⁵⁹ in presence of acetic acid or pyridine affording the monoacetate **116a** or 3α -succinoxy-12-ketocholanic acid **116b**. Upon chromic acid oxidation followed by alkaline treatment, ketone **118** was easily obtained (Scheme 25). The bromination of **117a-117b** with bromine at 65 - 75° in glacial acetic acid

solution gave a mixture of 11 α - and 11 β - bromo derivatives (*Scheme 26*).¹⁵⁷⁻¹⁶¹ Stefanovic and co-workers¹⁶² found that when bromination in presence of HBr was carried out at room temperature (48-72 h) instead at 75°, the pure crystalline equatorial 11 α -bromo-derivative **119a** was obtained almost

Table 11. ¹H NMR Chemical Shifts of Estra-3-methoxy-1,3,5(10)-trien-11-one Derivatives

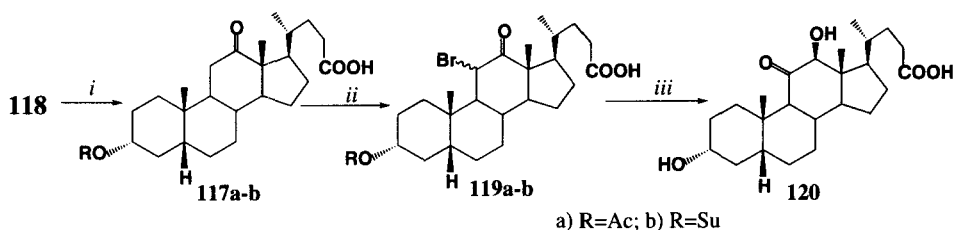
Compd.	substituents			Chemical Shifts (ppm)			ref.
	R ¹	R ²	R ³	C ₁ -H	C ₂ -H	C ₄ -H	
108	=H ₂	CH ₃	a-H	7.33	6.81	6.63	154
75c		CH ₃	a-H	7.20	6.68	6.56	121
109	=O	CH ₃	a-H	7.26	6.82	6.64	143
77c	β -OCH ₂ Ph	C ₂ H ₅	a-H	7.26	6.74	6.60	129
76j	=O	C ₂ H ₅	a-H	7.28	6.67	6.64	139
110	=H ₂	CH ₃	a-OH	7.35	6.77	6.62	154
111	=O	CH ₃	a-OH	7.33	6.97	6.83	143
112	β -OCH ₂ Ph	C ₂ H ₅	b-H	6.89	6.66	6.65	139
113	=O	CH ₃	b-H	6.72	6.72	6.72	143
75e		CH ₃	b-H	6.48-6.92	6.48-6.92	6.48-6.92	121
114	β -OH	C ₂ H ₅	b-OH	6.67	6.72	6.70	139
77g	β -OCH ₂ Ph	C ₂ H ₅	b-OH	6.68	6.74	6.60	139
79c	=O	CH ₃	b-OH	6.80	6.80	6.90	143
96b	=H ₂	CH ₃	b-OH	6.75	6.75	6.75	154



Reagents and Results: (i) Ac₂O, AcOH, reflux, 3 h, **116a**; succinic anhydride, pyridine, rt., overnight, **116b**: 91%; (ii) chromic acid, AcOH, rt., 25 min., **117a**; CrO₃, AcOH, H₂O, rt., 5 h, **117b**; (iii) **117a**, KOH, H₂O, reflux, 3 h, overall yield **115-118**: 74%; **117b**, alkaline solution, reflux, 3 h, overall yield **115-118**: 90%.

Scheme 25

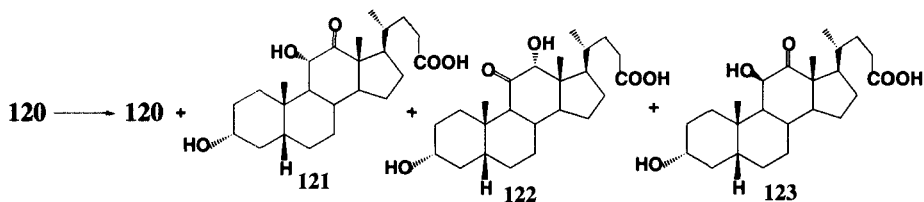
quantitatively. Application of the same bromination on sapogenins also resulted in the formation of 11 α -bromo-derivatives.¹⁶³ The 11-bromo-derivatives were then refluxed for two hours with a solution of sodium hydroxide in aqueous methanol to effect the rearrangement of the intermediate 11-hydroxy-12-ketone into the 12 β -hydroxy-11-ketone (Marker-Lawson acid, **120**).^{157, 161, 162} Gallagher¹⁶⁴ showed



Reagents and Results: (i) Ac₂O, AcOH, reflux, 1.5h, **117a**: 77%; succinic anhydride, pyridine, reflux, 1 h, **117b**: 83%; (ii) Br₂, AcOH, HBr, 25°, 48-72h, **119a** (11 α -): 100%; Br₂, AcOH, 65°, **119b** (11 α - + 11 β -): 98%; (iii) **119a**, NaOH, MeOH, reflux, 2 h, **120**: 79%; **119b**, NaOH, MeOH, reflux, 2 h, **120**: 77%.

Scheme 26

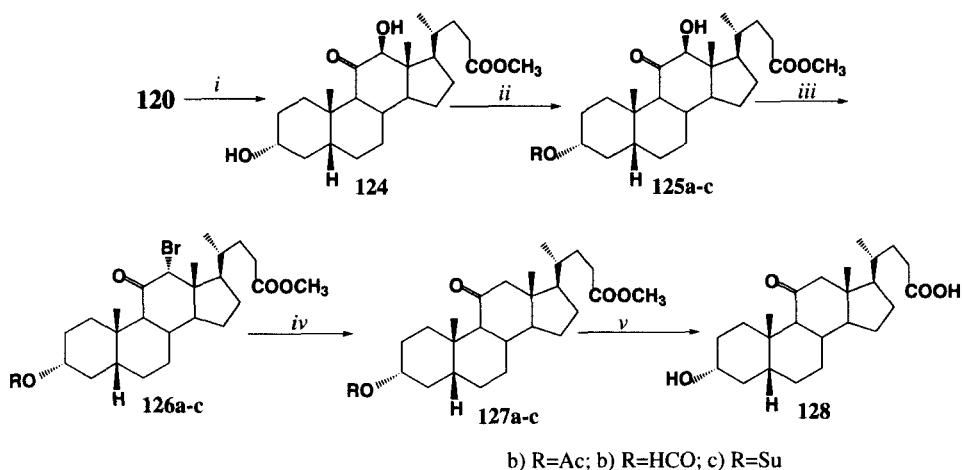
that all four possible ring C ketols were formed. The predominant isomer (60%) was the 12 β -hydroxy-11-keto acid **120**, but a substantial amount (33%) of the isomeric 11 α -hydroxy-12-keto acid contaminated with trace amounts of the other two isomers, was also present. The alkaline isomerization of 3 α ,12 β -dihydroxy-11-ketocholanic acid also gave the same four possible ketols (*Scheme 27*).¹⁶⁵ Monoacetylation of ester **124** was first described to proceed in a poor yield.¹⁶⁶ Later studies



Reagents and Results: 5% NaOH, reflux, 2 h, **120**: 63%; **121**: 30%; **122**: 1.1%; **123**: trace.

Scheme 27

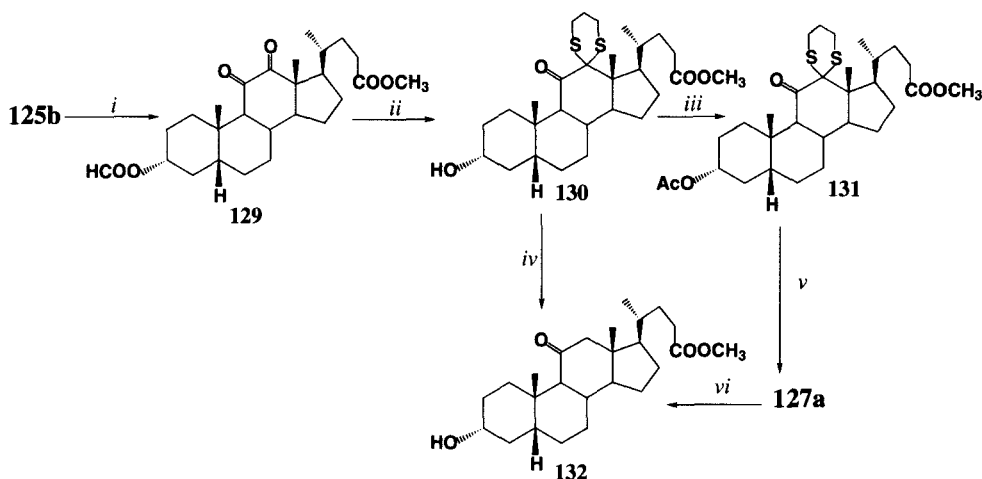
showed that this reaction may be effected readily in 80% yield by simple dissolution of the ester **124** in a four-fold excess of acetic anhydride (*Scheme 28*).¹⁶² The selective acylation of the ester **124** could be carried out in good yield in 90% formic acid.¹⁶⁶ The dimethyl ester of 3 α -succinoxy-12 β -hydroxy-11-ketocholanic acid **125c** was converted to the corresponding bromoketone **126c** in 50-70% yield with aid of phosphorus tribromide.¹⁶⁵ The inversion of C-12 configuration was not made on a suitable derivative with methyl 3 α -acetoxy-12 α -bromo-11-ketocholanoate **126a**^{18a} until 1967.¹⁶² Conversion of monoformate **125b** to the corresponding bromoketone monoformate **12b** was never accomplished in yields greater than 48% despite much experimentation.¹⁶⁶ In a variant of the above procedure, Hershberg¹⁶¹ converted 3 α -acetoxy-12 β -hydroxy-11-keto-24,24-diphenylcholene to the corresponding bromoketone in only 47% crude yield. In many experiments involving this procedure, a phosphorus-containing substance was isolated, the empirical formula of which corresponded to that of the 12-phosphite ester. Reduction of the bromoketones with Zn/HOAc led to the desired 11-ketone steroids (**127a** and **127c**).^{165, 18a}



Reagents and Results: (i) HCl, MeOH, rt., 2 h, **124**: 95%; (ii) Ac₂O, pyridine, 45 min., **125a**: 80%; HCOOH, 65°, 3 h, **125b**: 92%; succinic anhydride, pyridine, rt., overnight, **125c**: 91%; (iii) PBr₃, CH₂Cl₂, overnight, rt., **126a, b, c**: 70, 47, 50-60%; (iv) Zn, HOAc, reflux, 20 min., **127a**: 88%; Zn, HOAc, reflux, 1 h, **127c**: 98%; (v) 5N NaOH, MeOH, reflux, 30 min., **128**: 94%.

Scheme 28

An alternative way of elimination 11 β -OH in compound **125** was reported by Archer.¹⁶⁶ The pure ketol ester **125b** was first oxidized with chromic oxide to give the diketofornate **129** in 81% yield (Scheme 29). When the diketofornate **129** was treated with trimethylenedithiol in methanol

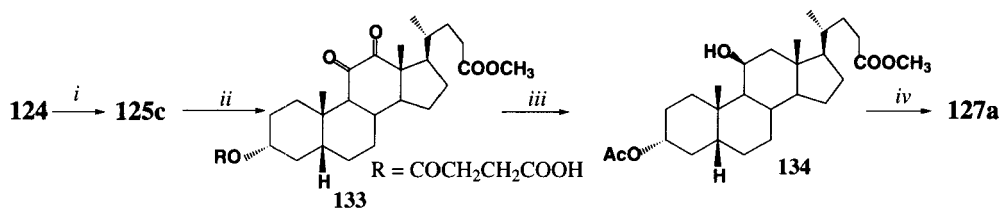


Reagents and Results: (i) CrO₃, HOAc, rt., 2 h, **129**: 81%; (ii) HCl, MeOH, trimethylene dithiol, rt., 2 h, **130**: 82%; (iii) Ac₂O, pyridine, rt., overnight, **131**: 88%; (iv) Raney nickel, MeOH, rt., 6 h, **132**: 84%; (v) Raney nickel, MeOH, rt., 6 h, **127a**: 95%; (vi) 5N KOH, reflux, 1 h, **132**: 93%.

Scheme 29

solution with hydrogen chloride gas as the condensing agent, the major product was the deformedylated product **130**. On subsequent acetylation, diketoacetate **131** was obtained. The Raney nickel desulfuration of **131** was best carried out at room temperature for a period of several hours. In this way, the conversion to pure 3α -acetoxy-11-keto-cholanate **127a** and 3α -hydroxy-11-ketone-cholanate **132** was accomplished in 95% yield and 93% yield, respectively.

Under the conditions of the Wolff-Kishner reaction, the 12-keto group also could be reduced to the methylene group (*Scheme 30*).¹⁶⁷ The non-crystalline reaction product was converted into acetylated methyl ester **134**. The enolization could be anticipated. The chromic acid oxidation of ester **134** finally afforded **127a**.

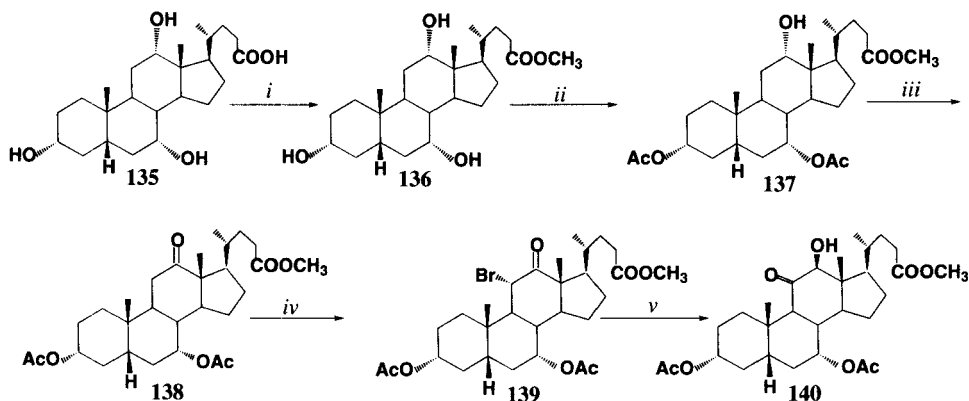


Reagents and Results: (i) succinic anhydride, pyridine, rt., 3 days, **125c**: 67%; (ii) CrO_3 , HOAc, rt., 1 h, **133**: 29%; (iii) hydrazine, EtOH, 200° , 6 h; diazomethane; Ac_2O , pyridine; **134**: 10%; (iv) CrO_3 , HOAc, rt., 17 h, **127a**: 91%.

Scheme 30

11-Oxocholeic Acid Derivatives

The application of Marker and Lawson's method to cholic acid derivatives was first reported by Yanuka and Halperin.¹⁶⁸ Initially, cholic acid (**135**, *Scheme 31*) was methylated in methanol and acetyl chloride, then selectively acetylated at the 3α - and 7α - position^{169, 170} to give the $3\alpha, 7\alpha$ -diacetoxy-12 α -hydroxy compound **137**.



Reagents and Results: (i) acetyl chloride, MeOH, rt., 2 h, **136**: 96%; (ii) Ac_2O , pyridine, benzene, rt., 2 h, **137**: 86%; (iii) K_2CrO_4 , AcOH, rt., 15 h, **138**: 94%; (iv) BF_3 , Br_2 , rt., 5 days, **139**: 95-97%; (v) KOH; diazomethane; Ac_2O , pyridine, rt., 18 h, **140**: 41%.

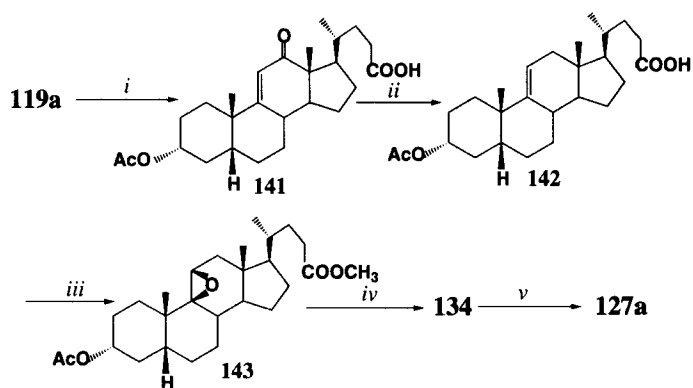
Scheme 31

Oxidation of this product with potassium chromate^{169, 170} afforded methyl 3 α ,7 α -diacetoxy-12-oxo-5 β -cholan-24-oate **138**. Jones' oxidation can also give satisfactory results.¹⁷¹ Exposure of **138** to the action of bromine at 70° in the presence of hydrobromic acid as a catalyst produced a complex mixture, with a low yield of bromo ketone **139**.¹⁶⁸ To improve the reaction HBr was substituted by BF₃, a strong Lewis acid, known to be very effective as a catalyst in bromination reactions.¹⁷² The desired 11 α -bromo ketone **139** was obtained in a high yield of 95-97%. Only minor amounts of the 11 β -bromo epimer could be detected. Conversion of this bromoketone **139** to the tautomerized ketol followed by treatment with diazomethane and acetic anhydride gave **140**.¹⁷³

2. Epoxidation

11-Oxo-deoxycholic Acid Derivatives

A mixture of 3 α -hydroxycholenic acids with double bonds at the C₉-C₁₁ and C₁₁-C₁₂ positions could be synthesized from deoxycholic acid.¹⁷⁴⁻¹⁷⁶ The introduction of C₉-C₁₁ double bond could be accomplished directly by dehalogenation of **119a** with sodium ethylate (*Scheme 32*).¹⁷⁴



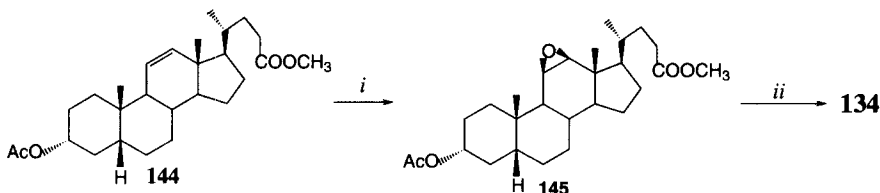
Reagents and Results: (i) sodium ethoxide, EtOH, reflux, 2 h, **141**: 77%; (ii) carbazide hydrochloride, sodium ethoxide, 180°, 15 h, **142**; (iii) HCl, MeOH; KMnO₄, HOAc, 0°, 2 h, **143**: 30%; (iv) H₂, PtO₂, HOAc, rt., 16 h, **134**: 93%; (v) CrO₃, HOAc, rt., 1 h, **127a**.

Scheme 32

Treatment of the acid with semicarbazide acetate gave a semicarbazone which was reduced by heating with sodium ethoxide in a sealed tube for fifteen hours at 180°. The application of perbenzoic acid to the C₉-C₁₁ double bond was shown by Chakravorty and Wallis¹⁷⁴ to yield an oxide mixed with some starting materials. The action of stronger oxidizing agent such as potassium permanganate, lead tetrapropionate, osmium tetroxide, hydrogen peroxide, and perbenzoic acid under a variety of conditions was investigated by Hick and co-workers.¹⁷⁷ The oxidation of methyl 3 α -acetoxy- $\Delta^{9,11}$ -cholenate with potassium permanganate resulted in formation of two isomeric oxides. The β -oxide was identified with an oxide prepared by the action of perbenzoic acid and the other oxide was not completely characterized. In the same reaction, Constantin and Sarett¹⁷⁸ found that the other "oxide" was inert to chromic acid and to perbenzoic acid and it was finally was identified as methyl $\Delta^{9,11}$ -3 α -

acetoxy-12-ketocholane rather than an epimeric oxide. The β -oxide readily underwent hydrogenolysis with platinum in acidic solution. Finally, oxidation of **134** with chromium trioxide in acetic acid gave methyl 3α -acetoxy-11-ketocholane **127a**.¹⁷⁸

Methyl 3α -acetoxy-11 β ,12 β -oxidocholane **145** was also prepared by potassium permanganate oxidation.^{177, 179} The action of hydrogen fluoride on this " β -oxide" followed by zinc reduction gave **134** (Scheme 33).

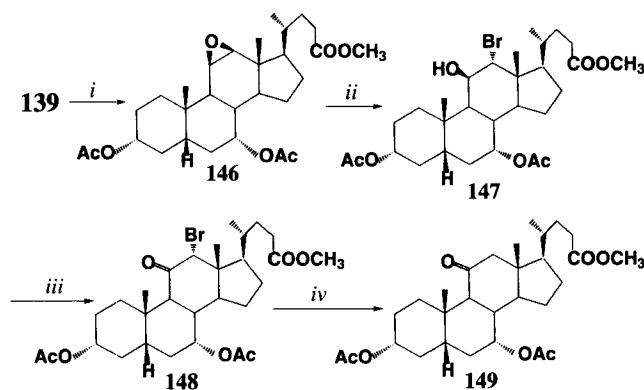


Reagents and Results: (i) KMnO_4 , HOAc, rt., 2 min., **145**: 61%; (ii) HF, -80° , 3 min.; Zn, HOAc, **134**

Scheme 33

11-Oxocholic Acid Derivatives.

The procedure for the preparation of the $3\alpha,7\alpha$ -diacetoxy-11-oxo-steroid **149** using methyl $3\alpha,7\alpha$ -diacetoxy-11 α -bromo-12-oxo-5 β -cholan-24-oate **139** as a starting material was adapted from that used¹⁸⁰ for the preparation of 7-deoxy- C_{11} -oxygenated-steroids (Scheme 34).¹⁸¹ The borohydride reduction of **139** in ethanol¹⁸⁰ or methanol¹⁸² under conditions similar to those utilized for 7-deoxy-steroids yielded a complex mixture. When pyridine was used as a solvent for borohydride reduction

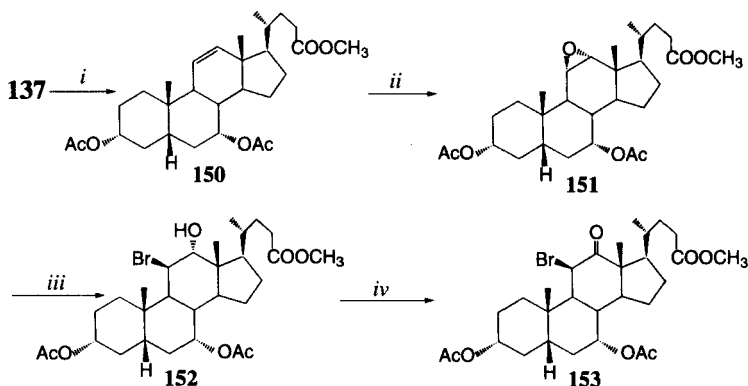


Reagents and Results: (i) NaBH_4 , NaOAc, pyridine, rt., 4 days, **146**: 32%; (ii) 48% HBr, HOAc, rt., 25 min., **147**: 48%; (iii) CrO_3 , AcOH, rt., 2.5 h, **148**: 94%; (iv) Zn, HOAc, reflux, 2 h, **149**: 92%.

Scheme 34

for 2 h, bromoketone **139** was not reduced. However, when the reaction time was prolonged and sodium acetate added, the epoxide **146** was formed directly. The epoxide was cleaved by hydrogen bromide followed by chromic acid oxidation to give ketone **148** which was reduced to **149**. During the

synthesis of 11 β -bromo ketone **153**, Yanuka and Halperin¹⁶⁸ applied perbenzoic acid to the C₁₁-C₁₂ double bond and obtained an α -oxide **151** (Scheme 35).



Reagents and Results: (i) POCl₃, pyridine, 37°, 24 h, **150**; (ii) PhCOOOH, CHCl₃, **137-151**: 18%; (iii) 48% HBr, acetone, **152**; (iv) CrO₃, HOAc, **153**: 80%.

Scheme 35

IV. CONCLUSION

11-Oxo steroids and their derivatives are very important intermediates in the synthesis of bioactive steroids such as cortisone, hydrocortisone, alphaxalone, desogestrel, 3-oxo desogestrel, mifepristone. The synthesis of 11-oxo steroids can be accomplished by the oxidation of 11-hydroxy steroids which can be obtained by microbiological transformation or partial synthesis. Epoxidation of $\Delta^{9(11)}$ steroids followed by hydrolysis can also produce 11-oxo steroids. Jones' oxidation on $\Delta^{9(11)}$ steroids can produce 9 β -hydroxy-11-oxo derivatives. Hydrolysis of 11 α -bromoketone can give 11-oxygenated steroids.

Acknowledgement.- The author wishes to thank Prof. J. R. Dias of this Department and referees for their numerous suggestions.

REFERENCES

1. M. G. Pomper, H. VanBrocklin, A. M. Thieme, R. D. Thomas, D. O. Kiesewetter, K. E. Carlson, C. J. Mathias, M. J. Welch and J. A. Katzenellenbogen, *J. Med. Chem.*, **33**, 3143 (1990).
2. a) R. D. Bindal, K. E. Carlson, G. C. A. Reiner and J. A. Katzenellenbogen, *J. Steroid Biochem.*, **28**, 361(1987); b) S. Sasson and J. A. Katzenellenbogen, *ibid.*, **33**, 859 (1989).
3. R. N. Hanson, E. Napolitano and R. Fiaschi, *J. Med. Chem.*, **33**, 3155 (1990).
4. R. Tedesco, R. Fiaschi and E. Napolitano, *J. Org. Chem.*, **60**, 5316 (1995).
5. S. Top, A. Vessieres and G. Jaouen, *Chem. Commun.*, 453 (1994).

6. E. Napolitano, R. Fiaschi, K. E. Carlson and J. A. Katzenellenbogen, *J. Med. Chem.*, **38**, 429 (1995).
7. a) F. Nique, P. Van De Velde, J. Bremaud, M. Hardy, D. Philibert and G. Teutsch, *J. Steroid Biochem. Mol. Biol.*, **50**, 609 (1994); b) A. Claussner, L. Nedelec, F. Nique, D. Philibert, G. Teutach and P. Van De Velde, *ibid.*, **41**, 609 (1992); c) T. Maudelonde, *Ann. Endocrinol.*, **52**, 387 (1991).
8. E. J. Bailey, H. Fazalerley, M. E. Hill, C. E. Newall, G. H. Phillipps, L. Stephenson and A. Tulley, *Chem. Commun.*, 106 (1970).
9. J. S. Baran, H. D. Lennon, S. E. Mares and E. F. Nutting, *Experientia*, **26**, 762 (1970).
10. A. J. Broek, A. I. A. Broess, M. J. Heuvel, H. P. Jongh, J. Leemhuis, K. H. Schonemann, J. Smits, J. D. Visser, N. P. Vliet and F. J. Zeelen, *Steroids*, **30**, 481 (1977).
11. A. J. Van den Broek, C. Van Bokhoven, P. M. J. Hobbelen and J. Lemhuis, *Recl. Trav. Chim. Pays-Bas*, **94**, 35 (1975).
12. a) S. J. Hopkins, *Drugs Today*, **18**, 361 (1982); b) S. L. Corson, *Am. J. Obstet. Gynecol.*, **168**, 1017 (1993); c) A. Phillips, D. W. Hahn, S. Klimek and J. L. McGuire, *Contraception*, **36**, 181 (1987); d) E. W. Bergink, A. D. Hanburger, E. De Jager and J. Van der Vies, *J. Steroid Biochem.*, **14**, 175 (1981); e) T. C. Van Soest, L. A. Van Dijck and F. J. Zeelen, *Recl. Trav. Chim. Pays-Bas*, **99**, 323 (1980).
13. a) D. J. Back, S. F. M. Grimmer, N. Shenoy and M. L. E. Orme, *Contraception*, **35**, 619 (1987); b) H. G. Hasenack, A. M. G. Bosch and K. Kaar, *ibid.*, **33**, 591 (1986).
14. L. Chen, S. Sheng, X. Fang, X. Yang, B. Chen, J. Shi, C. Liu and S. Peng, *Nanjing Yaoxueyuan Xuebao*, **17**, 282 (1986); *Chem. Abstr.*, **107**: 217911k (1987).
15. E. Ottow, S. Beier, W. Elger, D. A. Henderson, G. Neef and R. Wiechert, *Steroids*, **44**, 519 (1984).
16. a) G. Teutach and A. Belanger, *Tetrahedron Lett.*, 2051 (1979); b) G. Teutach, A. Belanger, D. Philibert and C. Tournemine, *Steroids*, **39**, 607 (1982); c) G. Teutsch, *Tetrahedron Lett.*, **23**, 4697 (1982); d) A. Belanger, D. Philibert and G. Teutsch, *Steroids*, **37**, 361 (1981).
17. E. C. Kendall, P. S. Hench, C. H. Slocumb and H. F. Polley, *Proc. Mayo Clin.*, **24**, 181 (1949).
18. a) R. B. Turner, V. R. Mattox, L. L. Engel, B. F. McKazie and E. C. Kendall, *J. Biol. Chem.*, **166**, 345 (1946); b) V. R. Mattox and E. C. Kendall, *ibid.*, **185**, 589 (1950).
19. E. O. Karow and D. N. Petsiavas, *Ind. Eng. Chem.*, **48**, 2213 (1956).
20. E. A. Weaver, H. E. Kenney and M. E. Wall, *Appl. Microbiol.*, **8**, 345 (1960).
21. I. Mateles and G. J. Fuld, *Antonie van Leeuwenhoek J. Microbiol. Serol.*, **27**, 33 (1961).

22. S. G. Knight, *Ann. N. Y. Acad. Sci.*, **139**, 8 (1966).
23. L. Tan and P. Falardeau, *J. Steroid Biochem.*, **1**, 221 (1970).
24. S. Kolhekar, D. T. Chaudhar, V. S. Salvi and B. B. Gaitonde, *Hind. Antibiot. Bull.*, **25**, 49 (1983).
25. S. Kolhekar and B. B. Gaitonde, *ibid.*, **26**, 41 (1984).
26. P. Somal and C. L. Chopra, *Appl. Microbiol. Biotechnol.*, **21**, 267 (1985).
27. V. Bihari, A. K. Joshi, R. C. Jaiswal, A. W. Khan and S. K. Basu, *J. Microb. Biotechnol.*, **1**, 41 (1986).
28. E. G. Ceen, J. P. R. Herrmann and P. Dunnill, *Enzyme Microb. Technol.*, **9**, 365 (1987).
29. V. Bihari, A. K. Joshi, A. W. Khan and S. K. Basu, *J. Microb. Biotechnol.*, **3**, 45 (1988).
30. T. K. Dutta, J. Datta and T. B. Samanta, *Biochem. Biophys. Res. Commun.*, **192**, 119 (1993).
31. J. Fried, R. W. Thoma, J. R. Gerke, J. E. Herz, M. N. Donin and D. Perlman, *J. Am. Chem. Soc.*, **74**, 3962 (1952).
32. I. Hiroshi, N. Atsushi and H. Masako, *Nippon Nogei-kagaku Kaishi*, **32**, 674 (1958).
33. G. Wix, M. Natonek and M. Koracs, *Acta Microbiol. Acad. Sci. Hung.*, **6**, 197 (1959).
34. L. Sallam, A. H. El-Refai and I. El-Kady, *Bull. Chem. Soc. Jpn*, **43**, 1239 (1970).
35. D. T. Nguyen, M. Mayer and M. M. Tanot, *C. R. Acad. Sci. Ser. D*, **272**, 2032 (1971).
36. A. F. Abdel-Fattah, L. A. R. Sallam, A. H. El-Refai and A. Zeinel-Abdin, *Pak. J. Biochem.*, **5**, 28 (1972).
37. L. A. R. Sallam, N. Naim, A. Zeinel, A. Badr and A. H. El-Refai, *Rev. Asoc. Bioquim. Argent.*, **41**, 221 (1976).
38. L. Sallam, N. Naim, A. Zeinel, A. Badr and A. El-Refai, *Rev. Latinoam. Microbiol.*, **19**, 151 (1977).
39. A. F. Abdel-Fattah, A. H. El-Refai, L. A. R. Sallam and M. A. Badawi, *Microbiol. Esp.*, **30**, 59 (1978).
40. F. Ergan, P. Atrat and P. Dhulster, *Z. Allg. Mikrobiol.*, **22**, 607 (1982).
41. M. N. Kim, F. Ergan, P. Dhulster, P. Atrat, G. Gelff and D. Thomas, *Biotechnol. Lett.*, **4**, 233 (1982).

42. H. E. Park and M. N. Kim, *Misaengmul Hakhoechi*, **27**, 70 (1989).
43. A. H. El-Refai, H. El-Monem, L. A. R. Sallam and I. A. El-kady, *Z. Allg. Mikrobiol.*, **10**, 183 (1970).
44. L. A. R. Sallam, A. H. El-Refai, S. Nada and A. F. Albdel-Fattal, *J. Gen. Appl. Microbiol.*, **19**, 155 (1973).
45. A. H. El-Refai and K. M. Ghanem, *Egypt J. Microbiol.*, **22**, 327 (1988).
46. D. H. Peterson and C. H. Murray, *J. Am. Chem. Soc.*, **74**, 1871 (1952).
- 47.a) D. H. Peterson, C. H. Murray, L. M. Eppstein, L. M. Reineke, A. Weintraub and P. D. Meister, *ibid.*, **74**, 5933 (1952); b) D. H. Peterson, C. H. Murray, L. M. Eppstein, L. M. Reineke, A. Weintraub and P. D. Meister, US Patent 2,602,769, 1952; [*Chem. Abstr.*, **46**: 8331f (1952)].
48. A. Ercoli and P. D. Ruggieri, *Gazz. Chim. Ital.*, **85**, 639 (1955).
49. O. Hanc, A. Capek, A. Simek, K. Macek and M. Tadra, *Czechoslov. Farm*, **6**, 119 (1957).
50. A. G. Timofeeva, A. S. Barmenkov and M. V. Fedotova, *Med. Orom. S. S. S. R.*, **11**, 23 (1957).
51. F. Rensser, H. J. Koepsell and G. M. Savage, *Appl. Microbiol.*, **9**, 346 (1961).
52. S. Alfred, H. Karl, O. Dicter, S. Sigfrid, S. Rudolf and Z. Joachim, *Z. Chem.*, **2**, 289 (1962).
53. L. A. R. Sallam, A. H. El-Refai and I. A. Elkady, *Z. Allg. Mikrobiol.*, **11**, 325 (1971).
54. E. J. Volker, *J. Chem. Educ.*, **54**, 65 (1977).
55. Y. B. Choi, S. K. Choi, H. S. Kim and Y. H. Park, *Sanap Misaengmul Hakhoechi*, **18**, 66 (1990).
56. M. H. Kim and M. N. Kim, *Misaengmul Hakhoechi*, **29**, 111 (1991).
57. M. H. Kim, J. H. Kim and M. N. Kim, *Misaengmul Hakhoechi*, **29**, 258 (1991).
58. J. Protiva, V. Schwarz, J. Martinkova and K. Syhora, *Folia Microbiol.*, **13**, 146 (1968).
59. J. J. Schneider, *J. Steroid Biochem.*, **5**, 9 (1974).
60. M. Morrin and O. P. Ward, *Biotechnol. Lett.*, **11**, 319 (1989).
61. K. Sonomoto, K. Nomura, A. Tanaka and S. Fukui, *Eur. J. Appl. Microbiol. Biotechnol.*, **16**, 57 (1982).
62. A. Capek and O. Hane, *Folia Microbiol.*, **7**, 121 (1962).

63. S. Makoto and T. Masako, *Chem. Pharm. Bull. Jpn*, **9**, 152 (1961).
64. G. M. Shull and D. A. Kita, *J. Am. Chem. Soc.*, **77**, 763 (1955).
65. K. Zetsche, *Naturwissenschaften*, **47**, 232 (1960).
66. A. H. El-Refai, I. A. R. Sallam and H. A. Geith, *Indian J. Exp. Biol.*, **13**, 400 (1975).
67. A. H. El-Refai, L. Sallam and H. Ceith, *Pak. J. Biochem.*, **8**, 13 (1975).
68. A. Bowers, J. S. Mills, C. Casas-Campillo and C. Djerassi, *J. Org. Chem.*, **27**, 361 (1962).
69. R. L. Pederson, J. A. Campbell, J. C. Babcock, S. H. Eppstein, H. C. Murray, A. Weintraub, R. C. Meeks, P. D. Meister, L. M. Reineke and D. H. Peterson, *J. Am. Chem. Soc.*, **78**, 1512 (1956).
70. a) L. L. Smith, G. Greenspan, R. Rees and T. Foell, *ibid.*, **88**, 3120 (1966); b) L. L. Smith, G. Greenspan, R. Rees and T. Foell, US Patent 3,231,589, 1966; *Chem. Abstr.*, **64**, 14926a (1966).
71. J. Flines, W. F. Van der Waard, W. J. Mijs and S. A. Szpilfogel, *Rec. Trav Chim. Pays-Bas*, **82**, 129 (1963).
72. Y. Y. Lin, M. Shibahara and L. L. Smith, *J. Org. Chem.*, **34**, 3530 (1969).
73. J. T. McCurdy and R. D. Garrett, *ibid.*, **33**, 660 (1968).
74. P. Karl, N. Gunter and E. Ulrich, Ger. Offen. DE 3,248,434, 1982; *Chem. Abstr.*, **101**: 149782t (1984).
75. H. W. Gao, X. D. Su, L. M. Zhou and Z. S. Li, *Org. Prep. Proced. Inter.*, **29**, 572 (1997).
76. H. W. Gao, X. D. Su and Z. S. Li, *Synth. Commun.*, **27**, 1981 (1997).
77. a) M. J. Van den Heuvel, C. W. Van Bokhoven, H. P. De Jong and F. J. Zeelen, *Recl Trav. Chim. Pays-Bas*, **107**, 331 (1988); b) M. J. Van den Heuvel, C. W. Van Bokhoven, H. P. DeJong and F. J. Zeelen, US Patent 3,998,813, 1976; *Chem. Abstr.*, **86**: 155860e (1977).
78. R. Joly, J. Warnant, J. Jolly and J. Mathieu, *Compt. Rend.*, **258**, 5669 (1964).
79. a) R. Joly, J. Warnant and J. Jolly, (Roussel-UCLAF) Fr. 1,375,078, 1964; *Chem. Abstr.*, **62**, 9201d (1965); b) R. Joly, J. Warnant and J. Jolly, (Roussel-UCLAF) Neth. Appl. 6,406,712, 1964; *Chem. Abstr.*, **63**, 18211g (1965); c) R. Joly, J. Warnant and J. Jolly, (Roussel-UCLAF) Ger. 1,229,526, 1966; *Chem. Abstr.*, **66**, 55647j (1967).
80. J. B. John and B. Seymour, (American Cyanamid Co.) US Patent 3,211,764, 1965; *Chem. Abstr.*, **64**, 12761a (1966).
81. M. Debono and R. M. Molloy, *Steroids*, **14**, 219 (1969).

82. R. Wang, W. Ding, Z. Zhou and X. Chen, *Zhongguo Yaowu Huaxue Zazhi*, **4**, 187 (1994). *Chem. Abstr.*, **122**, 240130a (1995).
83. A. L. Wilds and N. A. Nelson, *J. Am. Chem. Soc.*, **75**, 5366 (1953).
84. D. Burn and V. Petrow, *J. Chem. Soc.*, 364 (1962).
85. B. Aweryn and A. R. Daniewski, *Pol. J. Chem.*, **54**, 251 (1980).
86. R. Bucourt, A. Pierdet and J. Salmon, US Patent 3,906,096, 1975; *Chem. Abstr.*, **84**, 59873z (1976).
87. a) R. T. Gyar, Neth. Appl. 6,607,002, 1966; *Chem. Abstr.*, **66**, 65723v (1967); b) G. A. Hughes, H. Smith and D. Hartley, US Patent 3,624,110 (1968); c) G. A. Hughes, H. Smith and D. Hartley, US Patent 3,391,165 (1968).
88. J. J. Brown and S. Bernstein, *Steroids*, **8**, 87 (1966).
89. D. Hartley and G. Hughes, (Herchel Smith) Belg. 632,347, 1963; *Chem. Abstr.*, **61**, 1917a (1964).
90. Roussel-UCLAF Neth. Appl. 6,414,702, 1965; *Chem. Abstr.*, **63**, 18222g (1965).
91. a) M. Perelman and E. Farkas, (Eli Lilly & Co.) US Patent 3,086,027, 1963; *Chem. Abstr.*, **59**, 10182c (1963); b) L. F. Fieser and M. Fieser, "Reagents for Organic Synthesis", John Wiley and sons, Inc.: Vol. 1, p. 967, New York, 1967; c) M. Huebner, K. Ponsold, M. Oettel and R. Freund, *Arzneim-Forsch*, **30**, 401 (1980); d) M. Huebner, K. Ponsold and M. Oettel, Ger. Offen. 2,718,872, 1976; *Chem. Abstr.*, **88**, 121543e (1978); e) C. E. Cook, M. C. Wani, Y. W. Lee, J. R. Reel and D. Rector, WO 89 12,448, 1989; *Chem. Abstr.*, **112**, 198892q (1990); f) Q. Cheng, S. Li, L. Shao and R. Li, *Yiyao Gongye*, **16**, 399 (1985).
92. M. Perelman, E. Frkas, E. J. Fornefeld, R. J. Kray and R. T. Repala, *J. Am. Chem. Soc.*, **82**, 2402 (1960).
93. S. G. Levine and N. H. Eudy, *J. Org. Chem.*, **35**, 549 (1970).
94. Z. Wang and Z. Li, *Zhongguo Yiao Gongye Zazhi*, **22**, 227 (1991); *Chem. Abstr.*, **116**, 6803z (1992).
95. X. Su and Z. Li, *Dissertation of Beijing Medical University*, **1990**.
96. a) Steroid Research Group, Shanghai Institute of Org. Chem., *Academia Sinica Acta Chemica Sinica*, **33**, 139 (1975); b) R. W. Rees and S. Smith, US Patent 3,475,469 (1969).
97. G. Teichmueller, I. Lehmann, R. Gebuehr, E. Graupner, S. Krahrmer, G. Streibhardt and M. Wentzke, (Jenapharm G. m. b. H.) Ger. (East) DD 296,931, 1991; *Chem. Abstr.*, **116**, 174528g (1992).

98. G. Nomine, R. Bocourt and M. Vignau, US Patent 3,052,672, 1960; *Chem. Abstr.*, **57**, 16702f (1962).
99. P. Wieland and G. Anner, *Helv. Chim. Acta*, **50**, 1453 (1967).
100. B. Menzenbach, M. Huebner, R. Sahm and K. Ponsold, *Pharmazie*, **39**, 496 (1984).
101. J. J. Brown and S. Bernstein, *Steroids*, **1**, 113 (1963).
102. E. Ottow, S. Beier, W. Elger, D. A. Henderson, G. Neef and R. Wiechert, *ibid.*, **44**, 519 (1984).
103. G. Teichmueller, I. Lehmann, S. Krahmer, H. Henkel and A. Schmidt, (Jenapharm G. m. b. H.) Ger. (East) DD 298,243, 1988; *Chem. Abstr.*, **117**, 111893x (1992).
104. G. Teichmueller, I. Lehmann, R. Gebuehr, S. Krahmer, E. Wentzke and R. Zepfer, Ger. (East) DD 298,242, 1992; *Chem. Abstr.*, **117**, 8273w (1992).
105. G. Teichmueller, I. Lehmann, S. Krahmer, H. Henkel and Schmidt, A. Schmidt, (Jenapharm G. m. b. H.) Ger. (East) DD 298,244, 1988; *Chem. Abstr.*, **117**, 8725y (1992).
106. L. G. Liu, X. D. Su and Z. S. Li, *Synth. Commun.*, **25**, 3113 (1995).
107. P. Xia, Z. G. Zhang and S. Y. Yuan, *Chinese J. Org. Chem.*, **9**, 448 (1989).
108. L. G. Liu, T. Zhang and Z. S. Li, *Synth. Commun.*, **26**, 2999 (1996).
109. L. G. Liu, T. Zhang and Z. S. Li, *Tetrahedron*, **52**, 4495 (1996).
110. E. F. Whiting and A. W. Schneider, Ger. 1,111,179, 1959; *Chem. Abstr.*, **56**, 4829d (1962).
111. H. L. Herzog, M. J. Gentles, H. Marshall and E. B. Hershberg, *J. Am. Chem. Soc.*, **83**, 4073 (1961).
112. A. R. Hanze, US Patent 2,676,181, 1954; *Chem. Abstr.*, **49**, 5536c (1955).
113. R. E. Harmon, C. V. Zenarosa and S. K. Gupta, *J. Chem. Soc. D*, 537 (1969).
114. K. E. Pfitzner and J. G. Moffatt, *J. Am. Chem. Soc.*, **87**, 5670 (1965).
115. J. Schreiber and A. Eschenmoser, *Helv. Chim. Acta.*, **38**, 1529 (1955).
116. F. H. Westheimer and N. Nicolaides, *J. Am. Chem. Soc.*, **25**, 71 (1949).
117. K. E. Nicolaides and J. G. Moffatt, *ibid.*, **87**, 5661 (1965).
118. A. J. Mancuso, S. L. Huang and D. Swern, *J. Org. Chem.*, **43**, 2480 (1978).
119. G. Zweifel and H. C. Brown, *Org. React.*, **13**, 1-54 (1963).

120. R. V. Coomba, J. Koletar, R. Panna, H. Mah and E. Galantay, *J. Chem. Soc. Perkin Trans 1*, 2098 (1973).
121. J. Collins and J. Sjoval, *Aust. J. Chem.*, **36**, 339 (1983).
122. *Racemic compound*: H. Smith, *Brit. 1*, 1,28,044, 1968; *Chem. Abstr.*, **70**, 4388a (1969).
123. J. S. Li and Z. S. Li, *Yao Xue Xue Bao*, **20**, 187 (1985).
124. L. M. Zhou and Z. S. Li, *Dissertation of Beijing Medical University*, **1988**.
125. American Home Products Corp. *Brit. 1*, 214,506, 1970; *Chem. Abstr.*, **74**, 142175n (1971).
126. K. Ponsold and H. Kash, *Tetrahedron Lett.*, 4463 (1979).
127. L. Velluz and M. Legrand, *Angew. Chem.*, **77**, 844 (1965).
128. Y. H. Gong, "¹³C-NMR Shift of Natural Organic Products", p. 219-272, 1986.
129. H. W. Gao, X. D. Su and Z. S. Li, *Steroids*, **62**, 398-402 (1997).
130. G. A. Hughes and H. Smith, US Patent 3,787,395, 1972; *Chem. Abstr.*, **80**, 96223a (1974).
131. Z. S. Li and Y. Z. Ling, *Yao Xue Xue Bao*, **18**, 339 (1983). *Chem. Abstr.*, **99**, 212800e (1983).
132. S. Schwarz, S. Ring, G. Weber, G. Teichmueller, H.-J. Palme, C. Pfeiffer, B. Undetusch, B. Erhart and D. Grawe, *Tetrahedron*, **50**, 10709 (1994).
133. J. H. Fried, (Syntex Corp.) US Patent 3,408,372, 1967; *Chem. Abstr.*, **70**, 68646r (1969).
134. A. Pierdet and C. Bonne, *Ger. Offen.* 2,227,492, 1971; *Chem. Abstr.*, **78**, 72444f (1973).
135. A. Segaloff, R. B. Gabbard and A. Flores, *Steroids*, **35**, 335 (1980).
136. J. R. Hanson, P. B. Hitchcock, Y. Z. Ling and J. Y. Tan, *J. Chem. Res. (S)*, **47**, 46 (1985).
137. Z. S. Li and J. Y. Tan, *Yao Xue Xue Bao*, **18**, 501 (1983). *Chem. Abstr.*, **100**, 121429p (1984).
138. H. C. Brown and C. P. Garg, *J. Am. Chem. Soc.*, **83**, 2951 (1961).
139. H. Y. Wang and Z. S. Li, *Dissertation of Beijing Medical University*, **1991**.
140. H. Y. Wang and Z. S. Li, *Youji Huaxue*, **12**, 190 (1992). *Chem. Abstr.*, **117**, 48995q (1992).
141. H. Hasegawa, S. Nozoe and K. Tsuda, *Chem. Pharm. Bull. Jpn*, **11**, 1037 (1963).
142. H. Y. Wang and Z. S. Li, *Youji Huaxue*, **11**, 408 (1991). *Chem. Abstr.*, **115**, 280355q (1991).

143. H. Hasegawa and K. Tsuda, *Chem. Pharm. Bull. Jpn*, **12**, 473 (1964).
144. K. Tsuda, S. Nozoe and Y. Okada, *ibid.*, **11**, 1022 (1963).
145. a) C. D. Liang, J. S. Baran, N. L. Allinger and Y. Yuh, *Tetrahedron*, **32**, 2067 (1976); b) C. D. Liang, J. S. Baran, N. L. Allinger and Y. Yuh, US Patent 3,755,301, 1972; *Chem. Abstr.*, **79**, 137382b (1973).
146. W. K. Anderson and T. Veysoglu, *J. Org. Chem.*, **38**, 2267 (1973).
147. D. J. Collin and J. Sjovall, *Tetrahedron Lett.*, **7**, 629 (1979).
148. R. P. Stein, G. C. Buzby, G. H. Douglas and H. Smith, *ibid.*, 3603 (1967).
149. H. Y. Wang and Z. S. Li, *Youji Huaxue*, **12**, 51 (1992); *Chem. Abstr.*, **116**, 214764q (1992).
150. R. C. Cambie and T. D. R. Manning, *J. Chem. Soc; c*), 2603 (1968).
151. R. C. Cambie, V. F. Carlisle, C. J. L. Quesne and T. D. R. Manning, *ibid.*, 1234 (1969).
152. R. C. Cambie, V. F. Carlisle and T. D. R. Manning, *ibid.*, 1240 (1969).
153. R. C. Cambie and V. F. Carlisle, *ibid.*, 1706 (1970).
154. P. Aclinou, B. Gastambide, D. Deruaz, Y. Desage and R. Guilluy, *Tetrahedron*, **34**, 393 (1978).
155. T. G. Back, J. H.-L. Chau and M. Parvez, *Synthesis*, 162 (1995).
156. N. Cohen, B. L. Banner, W. F. Eichel, D. R. Parrish and G. Saucy, *J. Org. Chem.*, **40**, 681 (1975).
157. R. E. Marker and E. J. Lawson, *J. Am. Chem. Soc.*, **60**, 1334 (1938).
158. T. F. Gallagher and W. P. Long, *J. Biol. Chem.*, **162**, 521 (1946).
159. E. Schwenk, B. Riegel, R. B. Moffett and E. Stahl, *J. Am. Chem. Soc.*, **65**, 549 (1943).
160. O. Wintersteiner, M. Moore and K. Reinhardt, *J. Biol. Chem.*, **162**, 707 (1946).
161. E. B. Hershberg, L. Herzog, S. B. Coan, L. Weber and M. Jevnik, *J. Am. Chem. Soc.*, **74**, 2585 (1952).
162. M. Stefanovic, M. Gasic, J. Hranisavljevic and M. Dermanovic, *Tetrahedron Lett.*, 4799 (1967).
163. C. Djerassi, H. Martinez and G. Rosenkranz, *J. Org. Chem.*, **16**, 303 (1951).
164. T. F. Gallagher, *J. Biol. Chem.*, **162**, 539 (1946).

165. E. Borgstrom and T. F. Gallagher, *ibid.*, **177**, 951 (1949).
166. S. Archer, T. R. Lewis, C. M. Martini and M. Jackman, *J. Am. Chem. Soc.*, **76**, 4915 (1954).
167. O. Wintersteiner and M. Moore, *J. Biol. Chem.*, **162**, 725 (1946).
168. Y. Yanuka and G. Halperin, *J. Org. Chem.*, **38**, 2587 (1973).
169. L. F. Fiser, S. Rajagopalan, E. Wilson and M. Tishler, *J. Am. Chem. Soc.*, **73**, 4133 (1951).
170. L. F. Fiser and S. Rajagopalan, *ibid.*, **72**, 553 (1950).
171. A. Bowers, T. G. Halsall, E. R. H. Jones and A. J. Lemin, *J. Chem. Soc.*, 2555 (1953).
172. K. Takeda, T. Komeno and K. Igarashi, *Chem. Pharm. Bull. Jpn*, **4**, 343 (1956).
173. B. Nassim and J. R. Dias, *Org. Prep. Proced. Inter.*, **11**, 6 (1979).
174. P. N. Chakravorty and E. S. Wallis, *J. Am. Chem. Soc.*, **62**, 318 (1940).
175. E. M. Hicks, C. J. Berg and E. S. Wallis, *J. Biol. Chem.*, **162**, 633 (1946).
176. H. B. Alther and T. Reichstein, *Helv. Chim. Acta*, **25**, 805 (1942).
177. E. M. Hicks, C. J. Berg and E. S. Wallis, *J. Biol. Chem.*, **162**, 645 (1946).
178. J. M. Constantin and L. H. Sarett, *J. Am. Chem. Soc.*, **74**, 3908 (1952).
179. A. Furst and J. R. Scoton, *Helv. Chim. Acta*, **36**, 1410 (1953).
180. J. W. Conforth, J. M. Osbond and G. H. Phillips, *J. Chem. Soc.*, 907 (1954).
181. G. Halperin, *Steroids*, **33**, 295 (1979).
182. H. A. Walens and M. E. Wall, *J. Org. Chem.*, **26**, 1243 (1961).

(Received February 18, 1997; in revised form May 6, 1997)